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**VARIANTES GENÉTICAS ENVOLVIDAS NO  
METABOLISMO DO FOLATO: IMPACTO NA  
CARCINOGENESE DE CABEÇA E PESCOÇO**

Dissertação apresentada à Faculdade de  
Medicina de São José do Rio Preto para  
obtenção do Título de Mestre no Curso  
de Pós-graduação em Ciências da Saúde,  
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Ciências Correlatas.

Orientadora: Prof<sup>a</sup>. Dr<sup>a</sup>. Eny Maria Goloni-Bertollo

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*“Para adquirir conhecimento é preciso estudar, mas para adquirir sabedoria  
é preciso observar” Marilyn vos Savant*

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**Lista de abreviaturas e símbolos**

5-MTHF	5-metiltetrahydrofolato ( <i>5-methyltetrahydrofolate</i> )
5,10-MTHF	5,10-metilenotetrahydrofolato ( <i>5,10-methylenetetrahydrofolate</i> )
BHMT	Betaína-homocisteína metiltransferase ( <i>Betaine-homocysteine methyltransferase</i> )
Cb	Cobalamina
C $\beta$ S	Cistationina $\beta$ -sintase ( <i>Cystathionine <math>\beta</math>-synthase</i> )
CCP	Câncer de cabeça e pescoço
CEP	<i>Research Ethics Committee</i>
CH <sub>3</sub>	Metil ( <i>Methyl</i> )
CI 95%	<i>Confidence interval</i>
CNPq	Conselho Nacional de Desenvolvimento Científico e Tecnológico ( <i>National Council for Scientific and Technological Development</i> )
CONEP	Comitê Nacional de Pesquisa ( <i>National Research Commission</i> )
DHFR	Dihidrofolato redutase ( <i>Dihydrofolate reductase</i> )
DNA	Ácido desoxirribonucléico ( <i>Desoxirribonucleic acid</i> )
dTMP	Timidina monofosfato ( <i>Deoxythymidine monophosphate</i> )
dUMP	Deoxiuridina monofosfato ( <i>Deoxyuridine monophosphate</i> )
FAMERP	Faculdade de Medicina de São José do Rio Preto ( <i>São José do Rio Preto Medical School</i> )

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FAPESP	Fundação de Amparo à Pesquisa do Estado de São Paulo ( <i>São Paulo State Research Foundation</i> )
FUNFARME	Fundação Faculdade Regional de Medicina de São José do Rio Preto
HB	Hospital de Base
Hcy	Homocisteína ( <i>Homocysteine</i> )
HNC	<i>Head and neck câncer</i>
HNSCC	<i>Head and neck squamous cell carcinoma</i>
HPV	<i>Human Papiloma Virus</i>
INCA	Instituto Nacional do Câncer ( <i>Brazilian National Cancer Institute</i> )
L-MM-Coa mutase	L-metilmalonil coenzima A mutase ( <i>L-methylmalonyl coenzyme A mutase</i> )
M	Metástase á distância
MMA	Ácido metilmalônico ( <i>Methylmalonic acid</i> )
MTHF	Metilenotetrahidrofolato ( <i>Methylenetetrahydrofolate</i> )
MTHFD1	Metilenotetrahidrofolato desidrogenase 1 ( <i>Methylenetetrahydrofolate dehidrogenase 1</i> )
MTHFR	Metilenotetrahidrofolato redutase ( <i>Methylenetetrahydrofolate reductase</i> )
MTR	Metionina sintase ( <i>Methionine synthase</i> )

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MTRR	Metionina sintase redutase ( <i>Methionine synthase reductase</i> )
N	Envolvimento de linfonodos
OR	<i>Odds ratio</i>
PB	Pares de base
PCR	Reação em Cadeia da Polimerase ( <i>Polymerase chain reaction</i> )
PCR-RFLP	<i>Polymerase chain reaction-restriction fragment length polymorphism</i>
RFC1	Carregador de folato reduzido 1 ( <i>Reduced folate carrier 1</i> )
RNA	Ácido ribonucléico ( <i>Ribonucleic acid</i> )
SAH	S-adenosilhomocisteína ( <i>S-adenosylhomocysteine</i> )
SAM	S-adenosilmetionina ( <i>S-adenosylmethionine</i> )
SHMT	Serina hidroximetiltransferase ( <i>Serine Hydroxymethyltransferase</i> )
SISNEP	Sistema Nacional de Informação sobre Ética em Pesquisa ( <i>National Information System on Research Ethics</i> )
T	Tamanho do tumor
TC2	Transcobalamina 2 ( <i>Transcobalamin 2</i> )
THF	Tetrahydrofolato ( <i>Tetrahydrofolate</i> )
TNM	Classificação dos Tumores Malignos ( <i>TNM classification</i> )
TYS	Timidilato sintase ( <i>Thymidilate synthase</i> )
UICC	<i>International Union of Cancer Control</i>
UPGEM	Unidade de Pesquisa em Genética e Biologia Molecular ( <i>Genetics and Molecular Biology Research Unit</i> )

## RESUMO

**Introdução:** Câncer de cabeça e pescoço é um termo coletivo definido por bases anatômicas e topográficas para descrever tumores malignos do trato aerodigestivo superior. Esta região anatômica inclui a cavidade oral, faringe e laringe, tendo como principais fatores de risco o tabagismo e o etilismo. O tipo histológico mais representativo de todos os cânceres de cabeça e pescoço é o carcinoma espinocelular (HNSCC), com mais de 500 mil casos novos no mundo todos os anos. Deficiência de folato no organismo está associada ao aumento do risco de vários tipos de câncer e alterações neste metabolismo podem contribuir para o processo de carcinogênese por influenciar as reações de metilação do DNA e a estabilidade genômica. Polimorfismos em genes que codificam enzimas envolvidas no metabolismo do folato podem alterar a atividade enzimática e interferir nas concentrações de homocisteína, S-adenosilmetionina e outros produtos do metabolismo, importantes para a síntese de DNA e reações de metilação celular. **Objetivos:** Avaliar a influência dos polimorfismos *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G e *TC2* A67G em pacientes com carcinoma espinocelular de cabeça e pescoço e em indivíduos controle sem história da neoplasia, além de verificar a associação entre os polimorfismos e os sítios primários de ocorrência, extensão do tumor, comprometimento de linfonodos, e o prognóstico da doença. **Pacientes e Métodos:** Foram incluídos no estudo 694 indivíduos (240 pacientes com câncer de cabeça e pescoço e 454 controles). Foi feita análise molecular através de extração de DNA genômico de sangue periférico e as alterações genéticas foram investigadas por meio das técnicas de Reação em Cadeia da Polimerase (PCR) em tempo real e Análise de Polimorfismo de Comprimento de Fragmento de Restrição (PCR-RFLP). Os dados sócio-demográficos foram obtidos através do prontuário dos

pacientes e entrevista dos indivíduos controles. **Resultados:** Regressão logística múltipla mostrou que tabagismo, etilismo e idade superior a 42 anos foram preditores da doença ( $P < 0,05$ ). As distribuições genóticas estiveram em equilíbrio de Hardy-Weinberg em ambos os grupos em todos os polimorfismos estudados. Os genótipos *MTHFD1* 1958GA ou AA associados ao tabagismo ( $P=0,04$ ) e etilismo ( $P=0,03$ ) aumentaram o risco de carcinoma espinocelular de cabeça e pescoço. Estes mesmos genótipos estiveram presentes em maior proporção em pacientes com tumores em estadios mais avançados T3 e T4 ( $P=0,04$ ) e em pacientes com menor sobrevida ( $P=0,01$ ). O polimorfismo *TC2* C776G ( $P=0,03$ ) esteve presente em menor frequência em pacientes com idade superior a 52 anos e o polimorfismo *TC2* C776G ( $P=0,03$ ) em pacientes com idade entre 52-63 anos. O polimorfismo *TC2* C776G não foi relacionado ao risco da doença, porém esteve presente em alta proporção em pacientes que tiveram a faringe como sítio primário de ocorrência do tumor. **Conclusões:** São preditores para o câncer de cabeça e pescoço, independentemente da variável genética o uso de tabaco, álcool e idade superior a 42 anos. A presença do polimorfismo *MTHFD1* G1958A associado aos hábitos tabagista e etilista podem modular o risco para o desenvolvimento da doença.

**Palavras chave:** Polimorfismo genético, câncer de cabeça e pescoço, genes *MTHFD1*, *BHMT* e *TC*.



## ABSTRACT

**Introduction:** Head and neck cancer is a collective term defined by anatomical and topographical basis to describe malignant tumors of the upper aerodigestive tract. This anatomical region includes the oral cavity, pharynx and larynx, having as the main risk factors smoking and alcoholism. The most representative histologic type from head and neck cancer was squamous cell carcinoma (HNSCC), with more than 500,000 new cases worldwide every year. Folate deficiency is associated with increased risk of several types of cancer and alterations in folate metabolism may contribute to the process of carcinogenesis by influencing DNA methylation and genomic stability. Polymorphisms in genes encoding enzymes involved in this pathway may alter enzyme activity and consequently interfere in concentrations of homocysteine and S-adenosylmethionine that are important for DNA synthesis and cellular methylation reactions. **Objectives:** Investigate *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G and *TC2* A67G polymorphisms involved in folate metabolism on head and neck cancer risk, and the association between these polymorphisms with primary site, tumor extension, lymph node involvement and prognosis of the disease. **Patients and Methods:** Were included in the study 694 individuals (240 patients with head and neck cancer and 454 controls). Molecular analysis was made by genomic DNA from peripheral blood and genetic alterations were investigated by Polymerase Chain Reaction-restriction Fragment Length Polymorphism (PCR-RFLP) and Real Time-PCR. Socio-demographic data were obtained from patient's medical records and interview of the controls. **Results:** Multiple logistic regression showed that tobacco, alcohol and age over 42 years were predictors for the disease ( $P < 0.05$ ). Hardy-Weinberg equilibrium showed that the genotypic distributions were in equilibrium for both groups in all

polymorphisms studied. The *MTHFD1* 1958GA or AA genotypes associated with tobacco (P=0.04) and alcohol (P=0.03) consumption increase the risk for head and neck cancer (HNSCC). These same genotypes were found in higher proportion in patients with advanced stage tumors (P=0.04) and in patients with lower survival (P=0.01). *TC2* C776G polymorphism (P=0.03) were less frequent in patients with age over 52 years and *TC2* A67G polymorphism (P=0.04) were less frequent in patients with 52-63 years. *TC2* C776G polymorphism was not associated to HNC, however was present in higher proportion in patients with pharynx as primary site of tumor (P=0.02). **Conclusions:** Are predictors for head and neck cancer, regardless of the gene, tobacco and alcohol consumption and age over 42 years. The presence of *MTHFD1* G1958A polymorphism associated to tobacco and alcohol consumption may modulate the risk for disease development.

**Key words:** Genetic polymorphism; head and neck cancer; *MTHFD1*, *BHMT* and *TC2* genes.

# *1 INTRODUÇÃO*

## 1. INTRODUÇÃO

“Câncer de cabeça e pescoço” é um termo coletivo definido por bases anatômicas e topográficas para descrever tumores malignos do trato aerodigestivo superior. Esta região anatômica inclui a cavidade oral, faringe e laringe. Cerca de 40% dos cânceres de cabeça e pescoço ocorrem na cavidade oral, 15% na faringe e 25% na laringe. <sup>(1-3)</sup> É considerado o quinto tipo mais comum no mundo, está associado à baixa taxa de sobrevivência e alta taxa de mortalidade, quando diagnosticado em estágios avançados. <sup>(4)</sup> O tabagismo e o etilismo são os principais fatores de risco estabelecidos para o câncer de cabeça e pescoço. <sup>(4-12)</sup>

No Brasil ocupa o 5º lugar entre todas as neoplasias, com estimativas de 14.120 casos novos para o câncer de cavidade oral no ano de 2010, sendo 10.330 para o gênero masculino e 3.790 para o gênero feminino. <sup>(3)</sup> O comportamento desta neoplasia é bastante agressivo, apresentando metastatização cervical e contralateral precoce e, sobretudo na orofaringe, e em nódulos linfáticos cruzam a linha média da região cervical. <sup>(5)</sup>

O tipo histológico mais representativo de todos os cânceres de cabeça e pescoço é o carcinoma espinocelular (HNSCC), com mais de 500 mil casos novos no mundo todos os anos, <sup>(9)</sup> desenvolve-se a partir de um epitélio sujeito a um campo de cancerização, com mais agressividade na laringe. <sup>(1,13)</sup> Os fatores que promovem este campo de cancerização incluem exposições ambientais ao tabaco e álcool, infecções virais, especialmente com o vírus Epstein-Barr e Papiloma Vírus Humano dos subtipos 16 e 18 e deficiências ou desequilíbrios de vitaminas e micronutrientes, tais como ácido fólico, vitaminas A, C, E, zinco e selênio. <sup>(14-17)</sup>

Deficiência de folato no organismo está associada ao aumento do risco de câncer de cólon, <sup>(14,18)</sup> colorretal, <sup>(19-21)</sup> mama, <sup>(19-24)</sup> pulmão, <sup>(19-21,25)</sup> ovário, <sup>(19-21)</sup> esôfago, <sup>(26)</sup> colo de útero, <sup>(26)</sup> orofaringe, <sup>(27-29)</sup> estômago, <sup>(27-29)</sup> pâncreas, <sup>(27-29)</sup> rins, <sup>(30)</sup> e cabeça e pescoço. <sup>(31)</sup> Dietas ricas em frutas e legumes, que são fontes de ácido fólico e outros nutrientes antimutagênicos, são fortes proteções contra a maioria dos tipos de cânceres.

O folato possui importante papel na oncologia, principalmente a partir de sua ação na metilação do DNA e na síntese de purinas e pirimidinas. <sup>(27)</sup> Alterações genéticas ou de deficiência dessa vitamina foram relacionadas ao câncer em vários estudos, incluindo o de cabeça e pescoço. <sup>(18,21,22,26,27,31-35)</sup>

Existem três mecanismos pelos quais as alterações no metabolismo do folato podem contribuir com a carcinogênese: (1) hipometilação de DNA e subsequente ativação dos proto-oncogenes; <sup>(27,36)</sup> (2) erro de incorporação da uracila durante a síntese de DNA que leva à instabilidade genômica; <sup>(14,27,36)</sup> e (3) um aumento na desaminação de citosina nos sítios de metilação de DNA. <sup>(14,27)</sup>

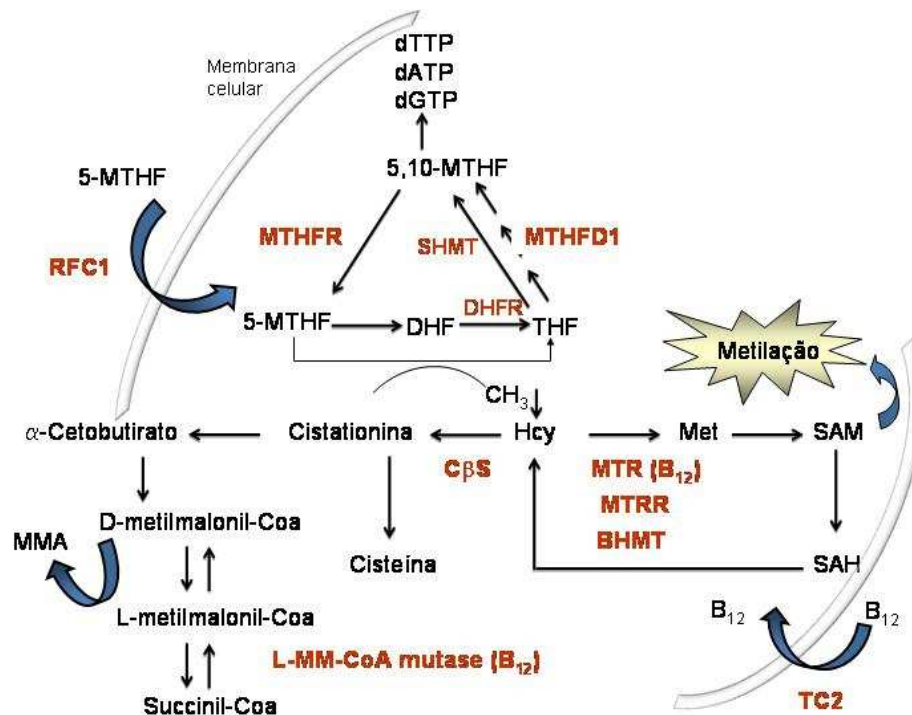
O folato está envolvido na formação de grupos metil (CH<sub>3</sub>) durante a interconversão de um carbono no metabolismo intermediário de S-adenosilmetionina (SAM), que serve como um doador de grupos metil nas reações de metilação celulares. <sup>(27,37-40)</sup> A metilação do DNA é a transferência de grupos metil para a posição 5 de resíduos de citosinas localizadas em dinucleotídeos citosina-guanina (CpG), por meio de reações catalisadas por proteínas denominadas DNA metiltransferases. <sup>(40,41)</sup> Esta modificação epigenética do DNA possui vários papéis funcionais, incluindo controle da expressão gênica, estabilidade da estrutura da cromatina e manutenção da estabilidade genômica. <sup>(27,38,41-46)</sup>

Níveis adequados de folato são essenciais para a biosíntese de purinas e pirimidinas, necessárias para a síntese e reparo do DNA. Portanto, alterações na via metabólica do folato estão associadas à redução na capacidade de reparo do DNA. A enzima Timidilato sintase catalisa a conversão de deoxiuridina (dUMP) a monofosfato de deoxitimidina (dTMP), utilizando 5,10 metilenotetrahidrofolato como doador de grupos metil. Em caso de deficiência de folato, o acúmulo de dUMP pode induzir a incorporação de uracila ao DNA ao invés de timina. As uracilas erroneamente incorporadas são removidas das fitas de DNA por enzimas da maquinaria de reparo, o que pode levar a quebras temporárias na molécula, posteriormente ligadas pela enzima DNA ligase. Entretanto, se a disponibilidade de folato é continuamente limitada, um ciclo de reparo descontrolado pode causar quebras frequêntes na molécula de DNA e danos cromossômicos, resultando em alteração celular maligna. <sup>(27,38,46-49)</sup> Outros nutrientes, como metionina, vitamina B<sub>6</sub> e vitamina B<sub>12</sub>, que interagem com genes envolvidos no metabolismo do folato, contribuem para a síntese adequada de DNA, e também podem influenciar o risco de desenvolvimento do câncer. <sup>(14,27,37,38,46)</sup>

Alterações em genes que codificam enzimas envolvidas na via do folato têm sido investigadas como fatores de risco para susceptibilidade ao câncer, uma vez que podem interferir nas concentrações de Hcy e SAM. <sup>(32-35,50-52)</sup>

Na Figura 1 são apresentadas as enzimas que participam do metabolismo do folato. A enzima Metilenotetrahidrofolato redutase (MTHFR) catalisa a conversão do 5,10 metilenotetrahidrofolato para 5-metiltetrahidrofolato (5-MTHFR), a principal forma circulante de folato, que atua como doador de grupos metil para a remetilação da homocisteína (Hcy) para metionina. Esta reação de remetilação é catalisada pela enzima Metionina sintase (MTR), que requer a vitamina B<sub>12</sub> (metilcobalamina) como cofator, e

resulta na formação de SAM. Participando também deste metabolismo, a enzima Cistationina  $\beta$ -sintase (C $\beta$ S), dependente de vitamina B<sub>6</sub>, desenvolve papel crucial no metabolismo do folato, convertendo a Hcy em cistationina na chamada via de transsulfuração. <sup>(53,54)</sup>



**Figura 1.** Esquema representando o metabolismo do folato com as principais enzimas envolvidas. BHMT = Betaína-homocisteína metiltransferase, C $\beta$ S = Cistationina  $\beta$ -sintase, dATP = Desoxiadenosina 5'-trifosfato, dGTP = Desoxiguanosina 5'-trifosfato, DHFR = Dihidrofolato redutase, dTTP = Desoxitimidina 5'-trifosfato, CH<sub>3</sub> = Metil, 5-MTHF = 5-metiltetrahydrofolato, 5,10-MTHF = 5,10-metilenotetrahydrofolato, Hcy = Homocisteína, L-MM-Coa mutase = L-metilmalonil coenzima A mutase, MMA = Ácido metilmalônico, MTHFD1 = Metilenotetrahydrofolato desidrogenase 1, MTHFR = Metilenotetrahydrofolato redutase, MTR = Metionina sintase, MTRR = Metionina sintase redutase, RFC1 = Carregador de folato reduzido 1, SAH = S-adenosil-homocisteína, SAM = S-adenosil-metionina, SHMT = Serina hidroximetiltransferase, TC2 = Transcobalamina 2 THF = Tetrahydrofolato.

### 1.1 Polimorfismos genéticos envolvidos no metabolismo do folato

O gene *Metileno-tetrahydrofolato desidrogenase 1 (MTHFD1)* codifica uma proteína trifuncional cistólica, que compreende 5,10-metileno-THF desidrogenase, 5,10-metenil-THF ciclohídroclase, e 10-formil-THF sintase. As enzimas metileno-THF desidrogenase e da metenil-THF ciclohídroclase, que residem no mesmo domínio da proteína, catalisam a oxidação do 5,10-metileno-THF a 5,10-metenil-THF, que é então convertido para 10-formil-THF. <sup>(55)</sup> Estas três reações sequenciais estão envolvidas na interconversão de derivados do carbono-1 do THF, que são substratos para a síntese de metionina, timidilato e purinas. <sup>(38,56)</sup>

O gene *MTHFD1* apresenta-se polimórfico no nucleotídeo 1958 (G→A) resultando na substituição de uma alanina por uma glicina no códon 653, localizado no domínio 10-formil-THF sintase da enzima. <sup>(38,57)</sup> Pacientes pediátricos com leucemia linfoblástica aguda, portadores dos alelos *MTHFD1* 1958A e timidilato sintase 2R (TS 2R), mostraram maior tempo de sobrevivência quando tratados com MTX. <sup>(52,57)</sup>

Também envolvido no metabolismo do folato, o gene Betaína-homocisteína metiltransferase (*BHMT*) apresenta-se polimórfico no nucleotídeo 742 (G→A), levando à substituição de arginina por glutamina na proteína produzida. <sup>(58)</sup> A avaliação do impacto deste polimorfismo nas propriedades funcionais da proteína BHMT resultante da variante polimórfica, não mostrou diferença na termoestabilidade e atividade catalítica em relação à enzima do tipo selvagem. <sup>(35)</sup> Xu e colaboradores (2008)<sup>(59)</sup> associaram o alelo polimórfico 742A com a redução dos casos de morte por câncer de mama. Por outro lado, Koushik e colaboradores (2006)<sup>(60)</sup> observaram um aumento do risco de desenvolvimento de câncer colorretal para portadores dos genótipos variantes



(742G/A e 742A/A) em relação aos portadores do genótipo homocigoto selvagem (742G/G).

Vitamina B<sub>12</sub> (cobalamina) e vitamina B<sub>6</sub> são nutrientes essenciais para o metabolismo do folato, uma vez que atuam como co-fatores de algumas enzimas envolvidas. A cobalamina (Cbl) possui um papel importante em 2 reações metabólicas: (1) conversão de L-metilmalonil-CoA a succinil-CoA e (2) remetilação da homocisteína para metionina. Assim, o transporte deste nutriente para a célula é importante para a manutenção do status de vitamina B<sub>12</sub> intracelular. <sup>(61,62)</sup>

Para a absorção celular de cobalamina é necessário que esta esteja ligada a uma proteína carregadora, a transcobalamina 2 (TC2). <sup>(61,62)</sup> A proteína TC2 é sintetizada no endotélio vascular da vilosidade intestinal e liga-se à vitamina B<sub>12</sub> livre no fluido intersticial. A proteína TC2 ligada à vitamina B<sub>12</sub> (complexo TC2-vitamina B<sub>12</sub>) passa, então, a microcirculação da vilosidade intestinal e, por meio da veia portal, alcança a circulação sistêmica. <sup>(63)</sup>

A presença de polimorfismos no gene *TC2* pode influenciar a quantidade de vitamina B<sub>12</sub> disponível no organismo. Estudos anteriores sugerem que um polimorfismo no nucleotídeo 776 (C→G) do gene *TC2* pode levar a alterações da proteína, influenciando sua afinidade e capacidade de transporte da cobalamina aos tecidos. <sup>(61,62)</sup> Concentração do complexo TC2-vitamina B<sub>12</sub> significativamente mais alta foi observada na presença do polimorfismo *TC2* C776G em homocigose para o alelo selvagem (776G/G). Além disso, concentrações médias de ácido metilmalônico (MMA), um indicador do *status* de vitamina B<sub>12</sub> <sup>(48)</sup> foram significativamente mais baixas na presença dos genótipos *TC2* 776C/C e 776C/G em relação ao genótipo 776G/G. <sup>(47)</sup>

O polimorfismo C776G leva a substituição de uma prolina por uma arginina no códon 259 (P259R) da transcobalamina. O polimorfismo C776G, também denominado de P259R (nomenclatura segundo alteração na proteína), altera a concentração plasmática da transcobalamina 2 livre (apo-TC2). Indivíduos portadores do genótipo PP apresentam maior concentração de apo-TC2 em comparação aos indivíduos com genótipos RR. Segundo Namour e colaboradores (2001),<sup>(64)</sup> a presença do alelo polimórfico em homozigose pode interferir na disponibilidade de Cbl intracelular e conseqüentemente no metabolismo da homocisteína.

O gene *TC2* também apresenta-se polimórfico no nucleotídeo 67 (A→G), localizado no éxon 2, resultando na substituição de uma isoleucina por uma valina no códon 23 (I23V).<sup>(61)</sup> Este polimorfismo foi associado com concentrações mais baixas da proteína produzida na presença do genótipo heterozigoto *TC2* 67A/G quando comparado ao genótipo tipo selvagem 67A/A,<sup>(61)</sup> entretanto, este genótipo também foi associado à proporção maior da proteína produzida ligada à vitamina B<sub>12</sub>.

Ainda não existem estudos que relacionam os polimorfismos do gene *TC2* ao câncer; porém estudo de Biselli e colaboradores (2008)<sup>(65)</sup> suportam evidências da relação entre esta variante genética em mães e a incidência de crianças portadoras de síndrome de Down. Este polimorfismo também foi associado à formação de aneurismas intracraniais.<sup>(66)</sup>

Dessa forma, o estudo de alterações em genes que participam do metabolismo do folato poderia auxiliar no esclarecimento dos processos que levam ao desenvolvimento de tumores, principalmente o câncer de cabeça e pescoço, colaborando na busca de novas estratégias de tratamento e prognóstico.

## 1.2 Objetivos

Considerando as evidências apresentadas, este estudo teve como objetivos:

1. Avaliar a associação dos polimorfismos *MTHFD1* G1958A, *BHMT* G742A, *TC2* A67G e *TC2* C776G no risco do câncer de cabeça e pescoço, em um estudo caso-controle.
2. Avaliar a associação dos polimorfismos com os hábitos tabagista e etilista, gênero e idade (fatores de risco) no desenvolvimento do câncer de cabeça e pescoço.
3. Verificar a associação entre os polimorfismos e os sítios primários de ocorrência, extensão do tumor, comprometimento de linfonodos e o prognóstico da doença.

*2 ARTIGOS CIENTÍFICOS*

## 2. Artigos Científicos

Os resultados referentes aos objetivos dessa dissertação estão apresentados na forma de artigo. No total estão apresentados 03 artigos submetidos para publicação.

### Artigo 1

**Título:** Head and neck cancer: impact of MTHFD1 G1958A polymorphism

**Periódico:** Revista da Associação Médica Brasileira (RAMB), aceito para publicação.

### Artigo 2

**Título:** *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G and *TC2* A67G polymorphisms and head and neck squamous cell carcinoma risk

**Periódico:** Molecular Biology Reports, submetido para publicação.

*ARTIGO CIENTÍFICO 1*

**Title: Head and neck cancer: impact of MTHFD1 G1958A polymorphism**

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## RESUMO

**INTRODUÇÃO:** Alterações no metabolismo do folato podem contribuir para o processo de carcinogênese por influenciar as reações de metilação do DNA e a estabilidade genômica. Polimorfismos em genes que codificam enzimas envolvidas nesse metabolismo podem interferir nas concentrações de homocisteína, S-adenosilmetionina e outros produtos importantes para a síntese de DNA e reações de metilação celular. **OBJETIVOS:** Investigar o polimorfismo *MTHFD1* G1958A envolvido no metabolismo do folato no risco para o câncer de cabeça e pescoço e verificar a associação entre esse polimorfismo com fatores de risco e características clínico-histopatológicas. **PACIENTES E MÉTODOS:** Estudo retrospectivo que avaliou o polimorfismo *MTHFD1* G1958A em 694 indivíduos (240 pacientes e 454 controles), por meio da técnica de Análise de Polimorfismo de Comprimento de Fragmento de Restrição. Para análise estatística foram utilizados os testes de regressão logística múltipla e qui-quadrado. **RESULTADOS:** Tabagismo e idade superior a 42 anos foram preditores da doença ( $P < 0,05$ ). Os genótipos *MTHFD1* 1958GA ou AA associados ao tabagismo ( $P = 0,04$ ) e etilismo ( $P = 0,03$ ) foram preditores da doença. Estes mesmos genótipos estão presentes em maior proporção em pacientes com tumores em estadios mais avançados ( $P = 0,04$ ) e em pacientes com menor sobrevida ( $P = 0,03$ ). **CONCLUSÃO:** A presença do polimorfismo *MTHFD1* G1958A associada aos hábitos tabagista e etilista aumenta o risco para desenvolvimento de câncer de cabeça e pescoço.

**Palavras-Chave:** Polimorfismo genético; Neoplasias de cabeça e pescoço; Gene *MTHFD1*.



**ABSTRACT**

**INTRODUCTION:** Alterations in folate metabolism may contribute to the process of carcinogenesis by influencing DNA methylation and genomic stability. Polymorphisms in genes encoding enzymes involved in this pathway may alter enzyme activity and consequently interfere in concentrations of homocysteine and S-adenosylmethionine that are important for DNA synthesis and cellular methylation reactions. **AIM:** Investigate *MTHFD1* G1958A polymorphism involved in folate metabolism on head and neck cancer risk and the association between this polymorphism with risk factors and clinical-histopathological parameters. **PATIENTS AND METHODS:** A retrospective study in *MTHFD1* G1958A polymorphism investigated in 694 individuals (240 patients and 454 controls) by Polymerase Chain Reaction-restriction Fragment Length Polymorphism. Multiple logistic regression and chi-square were used for the statistical analysis. **RESULTS:** Multivariable analysis showed that tobacco and age over 42 years were predictors for the disease ( $P<0.05$ ). The *MTHFD1* 1958GA or AA genotypes associated with tobacco ( $P=0.04$ ) and alcohol ( $P=0.03$ ) consumption were predictors of the disease. This polymorphism were more frequently in patients with advanced stage tumors ( $P=0.04$ ) and patients with lower survival ( $P=0.03$ ). **CONCLUSION:** The presence of *MTHFD1* G1958A polymorphism associated to tobacco and alcohol consumption increase the risk for head and neck cancer.

**Key words:** Genetic polymorphism; Head and neck cancer; *MTHFD1* gene.

## INTRODUÇÃO

As neoplasias de cabeça e pescoço são responsáveis por uma grande incidência de óbitos em todo o mundo, sendo considerado o 6º tipo mais comum<sup>1</sup>. A região anatômica afetada por esse tipo de tumor inclui principalmente a cavidade oral (40%), faringe (15%) e laringe (25%)<sup>2</sup>. Dados do Instituto Nacional do Câncer<sup>2</sup> mostraram que na população brasileira, há uma proporção de três casos no gênero masculino para cada caso no gênero feminino, localizados com maior incidência na cavidade oral.

O câncer de cabeça e pescoço tem como principais fatores de risco o tabagismo e o etilismo<sup>1</sup>. Infecções virais especialmente com o vírus Epstein-Barr e Papiloma Vírus Humano dos subtipos 16 e 18 e deficiências ou desequilíbrios de vitaminas e micronutrientes, tais como ácido fólico, vitaminas A, C, E, zinco e selênio também foram associados à ocorrência de neoplasias em câncer de cabeça e pescoço<sup>3-5</sup>.

O folato possui importante papel na oncologia, principalmente a partir de sua ação na metilação do DNA e na síntese de purinas e pirimidinas<sup>6</sup>. Alterações genéticas ou de deficiência dessa vitamina foram relacionadas ao câncer em vários estudos, incluindo o de cabeça e pescoço<sup>6-15</sup>.

O gene *Metilenotetrahidrofolato desidrogenase 1 (MTHFD1)* é responsável pela formação do 10-formil-THF, essencial para a síntese de DNA. Este apresenta-se polimórfico no nucleotídeo 1958 (G→A) resultando na substituição de uma alanina por uma glicina no códon 653, localizado no domínio 10-formil-THF sintase da enzima<sup>16</sup>. Se a disponibilidade de folato é continuamente limitada, um ciclo de reparo descontrolado pode causar quebras frequentes na molécula de DNA e danos cromossômicos, o que resulta em alteração celular maligna, contribuindo para o desenvolvimento do câncer<sup>6</sup>.

Poucos estudos avaliaram esse polimorfismo em câncer e os resultados são contraditórios. Kruszyna *et al.*<sup>17</sup> não encontraram diferenças estatísticas significantes na frequência genotípica e alélica do polimorfismo *MTHFD1* A1958G em pacientes com câncer de laringe. Matakidou *et al.*<sup>18</sup> e Chen *et al.*<sup>19</sup> não associaram o mesmo polimorfismo as neoplasias de pulmão e colorretal, respectivamente. Por outro lado, Li *et al.*<sup>20</sup> encontraram associação do polimorfismo *MTHFD1* G1958A com o câncer de mama.

Assim, os objetivos desse estudo foram investigar a frequência do polimorfismo *MTHFD1* G1958A em pacientes com carcinoma espinocelular de cabeça e pescoço e comparar com indivíduos sem história familiar da doença, e verificar se há associação entre esse polimorfismo e os fatores de riscos (tabagismo e etilismo) e características clínico-histopatológicas dos tumores (sítio primário de ocorrência, comprometimento de linfonodos e extensão tumoral).

## **PACIENTES E MÉTODOS**

A amostra desse estudo foi constituída de 694 indivíduos, 240 pacientes com câncer de cabeça e pescoço (grupo caso) e 454 indivíduos sem história de neoplasia (grupo controle), após obtenção do Termo de Consentimento Livre e Esclarecido (parecer 5566/2005 da Comissão de Ética em Pesquisa – CEP da Faculdade de Medicina de São José do Rio Preto – FAMERP).

Os pacientes foram incluídos no estudo após o diagnóstico histopatológico de carcinoma espinocelular realizado pelo Serviço de Otorrinolaringologia e Cirurgia de Cabeça e Pescoço do Hospital de Base de São José do Rio Preto/SP. Os tumores foram classificados de acordo com os parâmetros da *Union International Control Cancer* (IUCC), 2002 e *American Joint Committee for Cancer* (AJCC), 2002 em três critérios: tamanho do tumor (T), presença de linfonodos regionais comprometidos (N) e presença de metástase à distância (M). Quanto à localização anatômica do sítio primário do tumor, foram classificados em cavidade oral, faringe, laringe e sítio primário desconhecido<sup>21,22</sup>. O DNA das amostras de sangue foram provenientes do banco de amostras do laboratório e foram coletadas no período de março de 2000 a outubro de 2009.

O grupo controle consistiu em 454 indivíduos sem história de neoplasia e, por serem oriundos de um serviço de doação de sangue, são isentos de vinte tipos de doenças, conforme determina legislação brasileira (<http://www.hemonline.com.br/portarias/rdc153/indexframe.htm>). Os critérios para inclusão e exclusão foram, respectivamente, idade acima de 40 anos e história de neoplasia na família. Todos os participantes foram submetidos a uma entrevista para obtenção de variáveis, como idade, gênero e hábitos tabagista e etilista. Foram

considerados tabagistas indivíduos que consumiram cerca de 100 cigarros durante toda a vida e etilistas aqueles que ingeriram mais do que quatro drinques por semana<sup>23,24</sup>.

Para análise molecular, o DNA genômico foi extraído a partir de sangue periférico de acordo com a técnica de Miller *et al.*<sup>25</sup> com modificações. A técnica de Análise de Polimorfismo de Comprimento de Fragmento de Restrição (PCR-RFLP) foi utilizada para determinar os genótipos do polimorfismo *MTHFD1* G1958A. Os *primers* utilizados foram descritos por Hol *et al.*<sup>26</sup> (Sense: 5' – CACTCCAGTGTTTGTCCATG – 3'; Anti-sense: 5' – GCATCTTGAGAGCCCTGAC – 3'). A amplificação foi obtida com desnaturação inicial a 95°C por 5 minutos, seguida por 35 ciclos de 30 segundos para desnaturação do DNA a 95°C, 50 segundos de anelamento dos *primers* a 53°C e 90 segundos de extensão a 72°C. A extensão final foi realizada por 5 minutos a 72°C. O produto de 331pb foi submetido a digestão enzimática com a enzima *MspI* por 3 horas a 37°C. Os fragmentos de 166pb e 70pb foram gerados quando o alelo G esteve presente e o fragmento 266pb foi gerado quando o alelo A esteve presente.

A análise estatística foi realizada utilizando-se os programas computacionais Minitab/Windows - Versão 14.0, para avaliar os efeitos das variáveis analisadas em câncer de cabeça e pescoço e Bio Estat versão 3.0 para verificar se as distribuições enotípicas estavam em equilíbrio de Hardy-Weinberg. O teste de regressão logística múltipla foi utilizado para determinar o efeito das variáveis analisadas em câncer de cabeça e pescoço, que incluiu idade (referência: < 42 anos – idade em quartis), gênero (referência: feminino), hábito tabagista (referência: não fumantes) e hábito etilista (referência: não etilistas) e também para análise das variáveis clínico-histopatológicas. A classificação T foi dividida em tumores com pequena extensão (T1,T2) e com grande extensão (T3, T4). A classificação N foi dicotomizada em comprometimento de linfonodos negativo (N0), e positivo (N1, N2, N3). Os resultados foram apresentados em odds ratio (OR) e intervalo de confiança de 95% (IC – 95%). O nível de significância foi estabelecido em 5% ( $p \leq 0,05$ ). O método de Kaplan-Meier foi aplicado para avaliar a taxa de sobrevida considerando como ponto final da análise (*end point*) o período compreendido entre o diagnóstico da doença e o óbito.

## RESULTADOS

Os resultados do teste de regressão logística múltipla mostraram diferenças significantes entre pacientes e controles em relação às variáveis: tabagismo e idade superior a 42 anos ( $p < 0,05$ ) e, portanto, foram preditores da doença (Tabela 1).

O teste de Hardy-Weinberg mostrou que a distribuição genotípica estava em equilíbrio na amostra estudada (caso:  $X^2=0,7096$ ;  $P=0,3996$ , e controle:  $X^2=0,0707$ ;  $P=0,7903$ ). O polimorfismo *MTHFD1* G1958A não foi associado ao risco dessa doença. As frequências genotípicas *MTHFD1* 1958GG, GA e AA foram 35,83, 45,83, 18,34% respectivamente, para os casos, e 35,46, 48,68 e 15,86% respectivamente, para os controles. A frequência do alelo selvagem 1958G foi 0,59 e 0,6, e do alelo polimórfico 1958A foi 0,41 e 0,4 entre casos e controles, respectivamente.

Os resultados do teste de regressão logística múltipla para interação entre os fatores de risco e o polimorfismo *MTHFD1* G1958A mostraram que tabagismo (OR: 1,68; IC=95% 1,01-2,78;  $P=0,46$ ) e etilismo (OR:1,83; IC=95% 1,06-3,15;  $P=0,03$ ) associados aos genótipos *MTHFD1* 1958GA ou AA aumentou o risco para o desenvolvimento de câncer de cabeça e pescoço (Tabela 2).

Em relação aos parâmetros clínico-histopatológicos dos tumores, os resultados do teste de regressão logística múltipla mostraram associação com o estadiamento tumoral, no qual os genótipos *MTHFD1* 1958GA ou AA foram mais frequentes em indivíduos com estadio 3 e 4 ( $P=0,044$ ) (Tabela 3).

A média de sobrevida dos pacientes no período do estudo obtida pela estimativa de Kaplan-Meier foi de 82,57 meses para os pacientes com genótipo *MTHFD1* 1958GG e de 59,03 para os pacientes com genótipo *MTHFD1* 1958GA ou AA, conforme Figura 1 ( $P=0,031$ ).

## DISCUSSÃO

Os resultados mostraram que tabagismo e idade superior a 42 anos aumentam o risco para câncer de cabeça e pescoço, corroborando com dados da literatura, que confirmam que esse tipo de neoplasia é mais frequente a partir da 4ª década de vida<sup>1,27</sup> e em indivíduos tabagistas<sup>28-32</sup>.

O folato age como coenzima em várias reações celulares fundamentais e é necessário na divisão celular devido ao seu papel na biossíntese de purinas e pirimidinas, e, conseqüentemente, na formação do DNA e do RNA<sup>33</sup>.

O gene *Metileno-tetrahydrofolato desidrogenase 1 (MTHFD1)*, envolvido no metabolismo do folato, codifica uma proteína trifuncional cistólica que compreende 5,10-metileno-THF desidrogenase, 5,10-metenil-THF ciclohidrolase, e 10-formil-THF sintase. As enzimas metileno-THF desidrogenase e metenil-THF ciclohidrolase, localizadas no mesmo domínio da proteína, catalisam a oxidação do 5,10-metileno-THF a 5,10-metenil-THF, convertida para 10-formil-THF. Estas três reações seqüenciais estão envolvidas na interconversão de derivados do carbono-1 do THF, substratos para a síntese de metionina, timidilato e purinas<sup>19,34</sup>. O polimorfismo G1958A desse gene pode estar associado ao câncer devido a alterações na síntese de DNA e conseqüentemente, descontrole celular<sup>20</sup>.

No presente estudo foi observado que a distribuição genotípica está em equilíbrio, corroborando com a pesquisa de Kruszyna *et al.*<sup>17</sup>, que também não encontrou diferenças estatísticas significantes na freqüência genotípica e alélica do polimorfismo *MTHFD1* A1958G.

Em nosso estudo, o polimorfismo *MTHFD1* G1958A não foi associado ao risco de câncer de cabeça e pescoço, assim como os achados de Kruszyna *et al.*<sup>17</sup> em 131 pacientes com câncer de laringe e 250 indivíduos controles, Matakidou *et al.*<sup>18</sup> em 619 pacientes com câncer de pulmão e Chen *et al.*<sup>19</sup> em 274 pacientes com câncer colorretal e 461 indivíduos controles.

Entretanto, Li *et al.*<sup>20</sup> que avaliaram a 227 pacientes, mostraram que o genótipo polimórfico *MTHFD1* 1958AA ocorreu em maior proporção em pacientes com câncer de mama do que o genótipo selvagem *MTHFD1* 1958GG. Também foi encontrado no mesmo estudo associação entre uma maior freqüência de metilação em pacientes com câncer de mama e o genótipo polimórfico *MTHFD1* 1958AA.

Em nosso estudo houve uma interação significativa ente os genótipos *MTHFD1* 1958GA ou AA e hábitos tabagista e etilista, sugerindo que indivíduos com esses hábitos e genótipos GA ou AA possuem um risco maior no desenvolvimento do câncer de cabeça e pescoço. Não existem dados na literatura que comprovem essa associação.

A análise dos parâmetros clínico-histopatológicos confirmou que o tamanho do tumor T3 e T4 (avançado) foi mais freqüente em pacientes com genótipos GA ou AA. O estudo de Kruszyna *et al.*<sup>17</sup>, em análises de significância genotípica entre características do tumor, mostraram uma fraca associação dos genótipos *MTHFD1* e o tamanho do tumor.

A média de sobrevida dos pacientes durante o período do estudo obtida pela estimativa de Kaplan-Meier mostrou que pacientes com o genótipo selvagem *MTHFD1* 1958GG apresentaram uma média de sobrevida maior em relação aos pacientes com genótipos *MTHFD1* 1958GA ou AA (pelo menos um alelo polimórfico), confirmando uma associação entre a presença do alelo polimórfico e a diminuição do tempo médio de sobrevida. De acordo com levantamento bibliográfico realizado, esse é o primeiro estudo que avaliou a associação entre o tempo de sobrevida e a presença do polimorfismo.

## **CONCLUSÃO**

São preditores para o câncer de cabeça e pescoço, independentemente da variável genética o uso de tabaco e idade superior a 42 anos. A presença do polimorfismo *MTHFD1* G1958A associado aos hábitos tabagista e etilista aumentam o risco para o desenvolvimento do câncer de cabeça e pescoço. O polimorfismo é mais freqüente em tumores com estadios mais avançados da doença e em pacientes com menor prognóstico de vida. É importante corroborar por meio de outros estudos a influência do polimorfismo do gene *MTHFD1* e de outros genes envolvidos no metabolismo do folato na tumorigênese do câncer de cabeça e pescoço, para que seja determinada a etiologia e as correlações significativas com as características clínico-histopatológicas desses tumores.

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## **CONFLITO DE INTERESSE**

Não existem conflitos de interesse declarados em relação a este artigo.

**REFERÊNCIAS BIBLIOGRÁFICAS**

1. Argiris A, Karamouzis MV, Raben D, Ferris RL. Head and neck cancer. *Lancet* 2008; v.371.
2. INCA – Instituto Nacional de Câncer: [www.inca.gov.br](http://www.inca.gov.br), 2010.
3. Kane, MA. The role of folates in squamous cell carcinoma of the head and neck. *Cancer Detection and Prevention* 2006; 29, 46-53.
4. Lo AK, Lo KW, Tsao SW, Wong HL, Hui JW, To KF, *et al.* Epstein-Barr virus infection alters cellular signal cascades in human nasopharyngeal epithelial cells. *Neoplasia* 2006; 3:173-180.
5. Hennessey PT, Westra WH, Califano JA. Human papillomavirus and head and neck squamous cell carcinoma: recent evidence and clinical implications. *Dent Res* 2009; 88(4):300-6.
6. Linhart HG, Troen A, Bell GW, Cantu E, Chao W, Moran E, *et al.* Folate Deficiency Induces Genomic Uracil Misincorporation and Hypomethylation But Does Not Increase DNA Point Mutations. *Gastroenterology* 2009; 136:227–235.
7. Hsiung DT, Marsit CJ, Houseman EA, Eddy K, Furniss CS, McClean MD, *et al.* Global DNA methylation level in whole blood as a biomarker in head and neck squamous cell carcinoma. : *Cancer Epidemiol Biomarkers Prev.* 2007; 16:108-14.
8. Mu LN, Cao W, Zhang ZF, Yu SZ, Jiang QW, You NC, *et al.* Polymorphisms of 5,10-methylenetetrahydrofolate reductase (MTHFR), fruit and vegetable intake, and the risk of stomach. *Canc. Biomark.* 2007; 12: 61-75.
9. Ouerhani S, Oliveira E, Marrakchi R, Ben Slama MR, Sfaxi M, Ayed M, *et al.* Methylenetetrahydrofolate reductase and methionine synthase polymorphisms and risk of bladder cancer in a Tunisian population. *Cancer Genet Cytogenet* 2007; 176:48-53.
10. Pande M, Chen J, Amos CI, Lynch PM, Broaddus R, Frazier ML. Influence of Methylenetetrahydrofolate Reductase Gene Polymorphisms C677T and A1298C on Age-Associated Risk for Colorectal Cancer in a Caucasian Lynch Syndrome Population. *Cancer Epidemiol Biomarkers Prev* 2007; 16:1753-9.
11. Xu X, Gammon MD, Wetmur JG, Rao M, Gaudet MM, Teitelbaum SL, *et al.* A functional 19-base pair deletion polymorphism of dihydrofolate reductase (DHFR) and risk of breast cancer in multivitamin users. *Am J Clin Nutr* 2007; 85:1098–102.



12. Kalmbach RD, Choumenkovitch SF, Troen AP, Jacques PF, D'Agostino R, and Selhub J. A 19-Base Pair Deletion Polymorphism in Dihydrofolate Reductase Is Associated with Increased Unmetabolized Folic Acid in Plasma and Decreased Red Blood Cell Folate. *The Journal of Nutrition* 2008, 138: 2323–2327.
13. Ott N, Geddert H, Sarbia M. Polymorphisms in methionine synthase (A2756G) and cystathionine beta-synthase (844ins68) and susceptibility to carcinomas of the upper gastrointestinal tract. *J Cancer Res Clin Oncol* 2008; 134:405-10.
14. Garcia-Crespo D, Knock E, Jabado N, Rozen R. Intestinal Neoplasia Induced by Low Dietary Folate Is Associated with Altered Tumor Expression Profiles and Decreased Apoptosis in Mouse Normal Intestine. *The Journal of Nutrition*, 2009.
15. Langevin SM, Lin D, Matsuo K, Gao CM, Takezaki T, Stolzenberg-Solomon RZ, *et al.* Review and pooled analysis of studies on MTHFR C677T polymorphism and esophageal cancer. *Toxicology Letters* 2009; 184:73–80.
16. Krajcinovic M, Lemieux-Blanchard E, Chiasson S, Primeau M, Costea I, Moghrabi A. Role of polymorphisms in MTHFR and MTHFD1 genes in the outcome of childhood acute lymphoblastic leukemia. *The Pharmacogenomics Journal* 2004; 4, 66–72.
17. Kruszyna L, Lianeri M, Rydzanicz M, Gajecka M, Szyfter K, Jagodzinski PP. Polymorphic variants of folate metabolism genes and the risk of laryngeal cancer. *Mol Biol Rep* 2010; 37: 241-247.
18. Matakidou A, Galta R, Rudd MF, Webb EL, Bridle H, Eisen T, *et al.* Prognostic significance of folate metabolism polymorphisms for lung cancer. *British Journal of Cancer* 2007; 97: 247 – 252.
19. Chen J, Kyte C, Valcin M, Chan W, Wetmur JG, Selhub J, *et al.* Polymorphisms in the one-carbon metabolic pathway, plasma folate levels and colorectal cancer in a prospective study. *J. J Cancer* 2004; 110, 617–620.
20. Li SY, Rong M, Iacopetta B. Germ-line variants in methyl-group metabolism genes and susceptibility to DNA methylation in human breast cancer. *Oncol Rep.* 2006 Jan;15 :221-5.
21. Sobin LH, Wittelind CH. International union against cancer: TNM classification of malignant tumours. 6th edn. New York: Wiley; 2000.
22. Lee KJ. Essential Otolaryngology-Head & Neck Surgery. 8nd ed. New York: McGraw-Hill; 2003.

23. Kjaerhein K, Gaard M, Andersen A. The role of alcohol, tobacco, and dietary factors in upper aerogastric tract cancer: a prospective study of 10.900 Norwegian men. *Cancer Causes and Control* 1998; 9: 99-108.
24. Ahrendt SA, Chown JT, Yang SC, Wu L, Zhang MJ, Jen J, *et al.* Alcohol consumption and cigarette smoking increase the frequency of p53 mutations in non-small cell lung cancer. *Cancer Res* 2000; 3155-9.
25. Miller SA, Dikes DD e Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Research* 1988; 16: 1215.
26. Hol FA, Van der Put NM, Geurds MP, Heil SG, Trijbels FJ, Hamel BC, *et al.* Molecular genetic analysis of the gene encoding the trifunctional enzyme MTHFD (methylenetetrahydrofolate-dehydrogenase, methenyltetrahydrofolate-cyclohydrolase, formyltetrahydrofolate synthetase) in patients with neural tube defects. *Clinical Genetics* 1998; 2:119-125.
27. Werbrouck J, De Ruyck K, Duprez F, Van Eijkeren M, Rietzschel E, Bekaert S, *et al.* Single-nucleotide polymorphisms in DNA double-strand break repair genes: Association with head and neck cancer and interaction with tobacco use and alcohol consumption. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis* 2008; v.656, Issues 1-2, 30, p.74-81.
28. Psyrris A, DiMaio D. Human papillomavirus in cervical and head-and-neck cancer. *Nat Clin Pract Oncol.* 2006; 5: 24-31.
29. Guha N, Boffetta P, Wünsch Filho V, Eluf Neto J, Shangina O, Zaridze D *et al.* Oral health and risk of squamous cell carcinoma of the head and neck and esophagus: results of two multicentric case-control studies. *Am J Epidemiol.* 2007;166:1159–73.
30. Serefoglou Z, Yapijakis C, Nkenke E, Vairaktaris E. Genetic association of cytokine DNA polymorphisms with head and neck cancer. *Oral Oncology* 2008; 44:1093– 1099.
31. Yadav SS, Ruwali M, Shah PP, Mathur N, Singh RL, Pant MC, *et al.* Association of poor metabolizers of cytochrome P450 2C19 with head and neck cancer and poor treatment response. *Mutation Research* 2008; 644:31–37.
32. Leme CVD, Raposo LS, Ruiz MT, Biselli JM, Galbiatti ALS, Maniglia JV, *et al.* GSTM1 and GSTT1 genes analysis in head and neck cancer. *Rev Assoc Med Bras* 2010; 56(3): 299-303.

33. Krishnaswamy K, Nair KM, Importance of folate in human nutrition. *Br J Nutr* 2001; 85: S115-S24.
34. Stevens VL, McCullough ML, Pavluck AL, Talbot JT, Feigelson HS, Thun MJ *et al.* Association of Polymorphisms in One-Carbon Metabolism Genes and Postmenopausal Breast Cancer Incidence. *Cancer Epidemiol Biomarkers* 2007; 16(6).

**Tabela 1.** Distribuição demográfica, fatores de risco, genótipos e odds ratio (OR) para câncer de cabeça e pescoço.

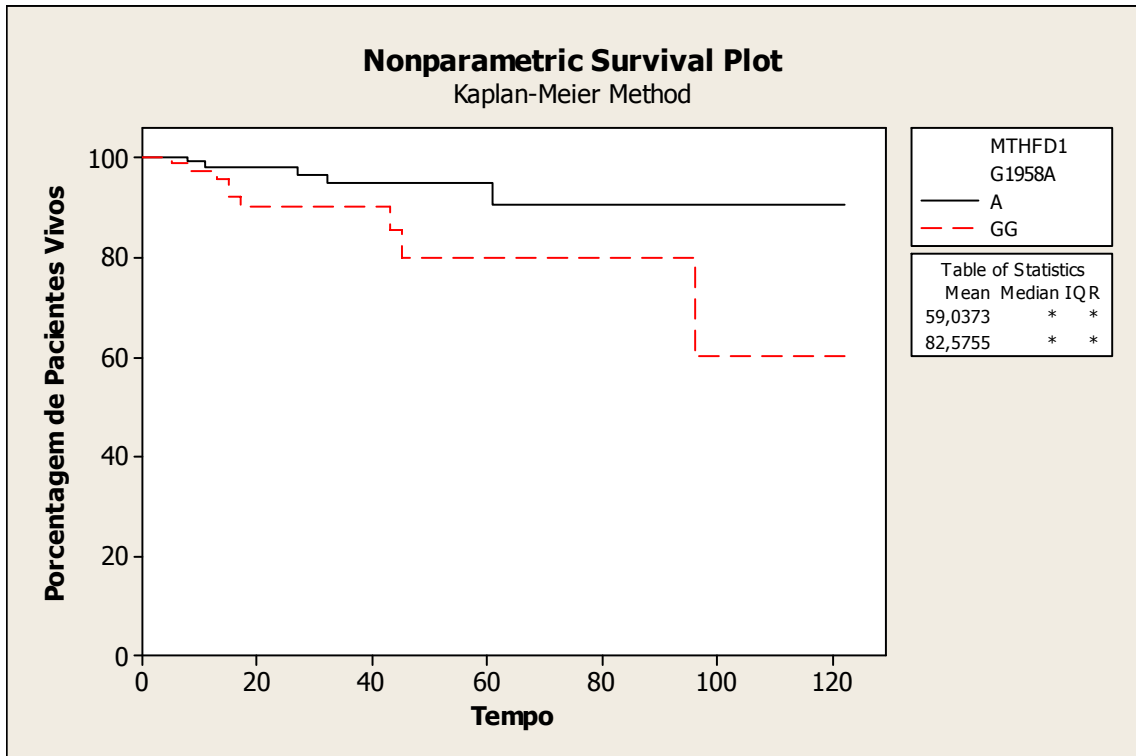
<b>Variáveis</b>	<b>Caso (%)</b>	<b>Controle (%)</b>	<b>OR (95%IC)</b>	<b>P value</b>
<b>Consumo de Tabaco</b>				
Não-fumantes	41 (17,08)	267 (58,81)	Referência	Referência
Fumantes	199 (82,92)	187 (41,19)	3,90 (2,46-6,20)	<b>P&lt;0,05</b>
<b>Consumo de Álcool</b>				
Não-etilistas	67 (27,92)	230 (50,66)	Referência	Referência
Etilistas	173 (72,08)	224 (49,34)	1,56 (0,99-2,48)	P=0,056
<b>Gênero</b>				
Feminino	29 (12,08)	129 (28,41)	Referência	Referência
Masculino	211 (87,92)	325 (71,59)	1,65 (0,95-2,86)	P=0,073
<b>Idade</b>				
<42 anos	8 (3,33)	177 (38,99)	Referência	Referência
42-51 anos	49 (20,42)	170 (37,44)	5,22 (2,53-10,77)	<b>P&lt;0,05</b>
52-63 anos	99 (41,25)	51 (11,23)	28,75 (13,51-61,18)	<b>P&lt;0,05</b>
>64 anos	84 (35)	56 (12,34)	24,51 (11,57-51,92)	<b>P&lt;0,05</b>
<b>Genótipo <i>MTHFD1</i></b>				
<b>G1958A</b>				
GG	86 (35,83)	161 (35,46)	Referência	Referência
GA	110 (45,84)	221 (48,68)	1,38 (0,91-2,10)	P=0,135
AA	44 (18,33)	72 (15,86)		

**Tabela 2.** Distribuição dos fatores de risco relacionados ao câncer de cabeça e pescoço e polimorfismo *MTHFD1* G1958A.

Variáveis	GG genótipo casos/controles	OR (95%IC)	GA e AA genótipos casos/controles	OR (95%IC)*	P value
<b>Idade</b>					
<42 anos	4/53	1,00 (ref)	6/123	0,38 (0,09-1,50)	P = 0,166
42-51 anos	19/64	1,00 (ref)	36/105	1,78 (0,83-3,80)	P = 0,136
52-63 anos	31/19	1,00 (ref)	60/32	2,02 (0,86-4,79)	P = 0,108
>64 anos	30/22	1,00 (ref)	54/34	1,31 (0,60-2,83)	P = 0,496
<b>Gênero</b>					
Feminino	10/42	1,00 (ref)	19/87	1,41(0,50-3,96)	P = 0,519
Masculino	76/135	1,00 (ref)	119/206	1,37 (0,85-2,19)	P = 0,192
<b>Tabaco</b>					
Não	23/97	1,00 (ref)	18/170	0,98 (0,45-2,14)	P = 0,964
Sim	63/64	1,00 (ref)	136/123	1,68 (1,01-2,78)	<b>P = 0,046</b>
<b>Álcool</b>					
Não	29/77	1,00 (ref)	38/153	0,95 (0,49-1,85)	P = 0,890
Sim	57/84	1,00 (ref)	116/140	1,83 (1,06-3,15)	<b>P = 0,030</b>

**Tabela 3.** Distribuição dos parâmetros clínico-histopatológicos e polimorfismo *MTHFD1* G1958A.

Parâmetros clínicos	GG genótipo casos (%)	OR (95%IC)	GA e AA genótipos casos (%)	OR (95%IC)*	P value
<b>Sítio Primário</b>					
Cavidade oral	35 (14,58)	1,00 (ref)	61 (25,42)	0,88 (0,51-1,53)	P = 0,659
Faringe	15 (6,25)	1,00 (ref)	36 (15)	1,42 (0,72-2,81)	P = 0,312
Laringe	28 (11,67)	1,00 (ref)	45 (18,75)	0,80 (0,45-1,43)	P = 0,454
<b>Tamanho do tumor</b>					
T1/T2	47 (19,58)	1,00 (ref)	107 (44,58)	1,00 (ref)	
T3/T4	37 (15,42)	1,00 (ref)	49 (20,42)	0,57 (0,32-0,98)	<b>P = 0,044</b>
<b>Envolvimento de linfonodos</b>					
Não	58 (24,17)	1,00 (ref)	111 (46,25)	1,00 (ref)	
Sim	26 (10,83)	1,00 (ref)	45 (18,75)	0,90 (0,50-1,62)	P = 0,721



**Figura 1.** Curva de sobrevivência não-paramétrica (Kaplan-Meier) dos pacientes com carcinoma espinocelular de cabeça e pescoço.

*ARTIGO CIENTÍFICO 2*



**Title: *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G and *TC2* A67G polymorphisms and head and neck squamous cell carcinoma risk**

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**ABSTRACT**

**INTRODUCTION:** Alterations in folate metabolism may contribute to the process of carcinogenesis by influencing DNA methylation and genomic stability. Polymorphisms in genes encoding enzymes involved in this pathway may alter enzyme activity and consequently interfere in concentrations of homocysteine and S-adenosylmethionine that are important for DNA synthesis and cellular methylation reactions. The objectives were to investigate *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G and *TC2* A67G polymorphisms involved in folate metabolism on head and neck cancer risk and the association between these polymorphisms with risk factors. **PATIENTS AND METHODS:** Polymorphisms were investigated in 762 individuals (272 patients and 490 controls) by Polymerase Chain Reaction-restriction Fragment Length Polymorphism (PCR-RFLP) and Real Time-PCR. Chi-square and Multiple logistic regression were used for the statistical analysis. **RESULTS:** Multiple logistic regression showed that tobacco and male gender were predictors for the disease ( $P < 0.05$ ). Hardy-Weinberg equilibrium showed that the genotypic distributions were in equilibrium for both groups in all polymorphisms studied. The *BHMT* 742GA or AA genotypes associated with tobacco consumption ( $P = 0.016$ ) increase the risk for head and neck squamous cell carcinoma (HNSCC). **CONCLUSION:** The present study suggests that *BHMT* 742GA associated to tobacco modulate HNSCC risk. However, further investigation of gene-gene interactions in folate metabolism and studies in different populations are needed to investigate polymorphisms and HNSCC risk.

**Key words:** Genetic polymorphism; Head and neck cancer; *MTHFD1*, *BHMT* and *TC2* genes.

## INTRODUCTION

Head and neck squamous cell carcinoma (HNSCC) is an aggressive malignant tumour type arising from the epithelial mucosal membranes of the upper-aerodigestive tract (oropharynx, hypopharynx and larynx) and the oral cavity [1,2]. HNSCC is the fifth most common cancer worldwide and is associated with low survival and high morbidity when diagnosed in advanced stage. Tobacco and alcohol consumption have been described as the most important risk factors associated with this carcinoma [3].

Folate is an essential nutrient which plays important roles in DNA synthesis and methylation [4-6]. Three main molecular mechanisms have been proposed: (1) a global decrease in DNA methylation, (2) increased uracil misincorporation during DNA replication, and (3) increased cytosine deamination at sites of DNA methylation. Folate metabolites are required for the conversion of homocysteine to methionine, which in the activated form of S-adenosyl-methionine (SAM) is required for DNA methylation. Folate deficiency can therefore decrease global DNA methylation, which is associated with genetic instability and tumor formation [7].

Low folate intakes have been positively associated with colon [8-11], breast [12-15], lung [12,13,16,17], colorectal [12,13,16,17], cervical [7,16], esophageal [7,16], pancreas [7,16], ovary [12,13,17] and head and neck cancer [18].

A polymorphism in methylenetetrahydrofolate dehydrogenase 1 (*MTHFD1*) gene (1958G>A) results in the substitution of a conserved arginine amino-acid by a glutamine at position 653 [19]. Despite the role of this enzyme in folate pathway, this polymorphism has been little explored in cancer [20,21].

Also involved in folate pathway, betaine-homocysteine methyltransferase (BHMT) is the enzyme, along with methionine synthase, which remethylates homocysteine (Hcy) to methionine [13,17,22,23]. The *BHMT* gene is polymorphic in the nucleotide 742, with a substitution of arginine for glutamine in the protein (G>A) [17,22,24,25]. There are few studies on the influence of this polymorphism on the development of cancer. Koushik *et al* (2006) [26] observed a relation between this polymorphism with colorectal cancer and Xu *et al* (2009) [27] with breast cancer.

A polymorphism in transcobalamin II (*TC2*) gene (776C>G) results in the substitution of a proline amino-acid by an arginine at codon 259 (P259R) [19,28]. Previous studies suggest that the C776G polymorphism in the *TC2* gene may affect transcobalamin binding affinity for Cbl and the ability to transport Cbl into tissues [28,29]. A different polymorphism in the *TC2* gene, the 67A>G transition is located in exon 2 and results in an isoleucine by valine replacement at codon 23 (I23V). There are no studies that have associated *TC2* polymorphisms and cancer; although Biselli *et al* (2008) [30] observed an association between this polymorphisms and maternal risk for Down syndrome, which etiology is related to abnormal folate metabolism. Afman *et al* (2002) [28] did not associate *TC2* C776G and A67G polymorphisms with neural tube defects risk.

Thus, the objectives of this study were to investigate *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G and *TC2* A67G polymorphisms involved in folate metabolism on head and neck cancer risk and the association between these polymorphisms with risk factors.

## PATIENTS AND METHODS

After approval by the National Ethics Committee (CONEP - 5566/2005; SISNEP 0976.0.140.000-05), the individuals who agreed to participate in the study signed an informed consent. A total of 762 individuals (272 patients with head and neck cancer and 490 controls) were included in the study. The diagnosis was made from pathological specimens after biopsy or total excision of the tumor. The inclusion criterion was squamous cell carcinoma tumor cell types and the exclusion criterion was patients previously treated for tumors.

The control group consisted of 490 Brazilian blood donors without a diagnosis of cancer according to government guidelines for donated blood that is tested for 20 related diseases (<http://www.hemonline.com.br/portarias/rdc153/indexframe.htm>) [31]. The inclusion criterion was age higher than 40 years and the exclusion criterion was a family history of cancer. Each eligible subject was interviewed to obtain data on gender, smoking habit, use of alcohol, and family history of cancer. The variables analyzed were gender and exposure to risk factors (tobacco and alcohol consumption). Individuals who had smoked more than 100 cigarettes in their lifetime were considered to be tobacco consumers and individuals who drank 4 doses of alcohol per week were considered to be alcohol consumers [32,33].

Genomic DNA was obtained from peripheral blood according to Miller *et al.* (1988) [34]. The *MTHFD1* G1958A and *TC2* C776G polymorphisms were investigated by polymerase chain reaction with restriction fragment length polymorphism (PCR-RFLP), according to Hol *et al.* (1998) [35] and Pietrzyk *et al.* (2003) [36], with some alterations. The primers and enzyme used were: sense: 5'-CAC TCC AGT GTT TGT CCA TG-3', anti-sense: 5'-GCA TCT TGA GAG CCC TGA C-3' and *MspI* for *MTHFD1* gene; and sense: 5'-CAT CAG AAC AGT GCG AGA GG-3', anti-sense: 5'-GTG CCA GAC AGT CTG GGA AG-3' and *ScrFI* for *TC2* gene. The *BHMT* G742A and *TC2* A67G polymorphisms were investigated by Allelic Discrimination (*Applied Biosystems*, USA) using TaqMan probes (TaqMan SNP Genotyping Assay C\_11646606\_20 and C\_25967461\_10, respectively) in Step One Plus™ Real-Time PCR System equipment (*Applied Biosystems*).

## STATISTICAL ANALYSIS

Multiple logistic regression was used to determine the interaction effect between the genetic polymorphisms and risk factors related to HNSCC. The model included gender (reference: female), tobacco consumption (reference: non-smokers) and alcohol consumption (reference: non-drinkers) using the Minitab for Windows computer program (Version 14.0). The Chi square test was used for to verify whether the genotypes frequencies were in Hardy-Weinberg equilibrium with BioEstat program. For deleterious alleles, the analysis was made using the criteria: risk 0 for no allele, risk 1 for one deleterious allele, risk 2 for two deleterious alleles, risk 3 for three deleterious alleles and risk 4 for four deleterious alleles or more.  $P < 0.05$  was considered statistically significant. Results are shown as odds ratio (OR) and 95% confidence intervals (95%CI). The Kaplan-Meier method was used to evaluate survival rates, considering as *end point* the period between the diagnosis of disease and obit.

## RESULTS

The results for comparison between groups showed that tobacco ( $P < 0.05$ ) and male gender ( $P < 0.05$ ) were predictors of the disease. Five hundred and eighty-seven (77.03%) participants were men (239 patients and 348 controls) and 175 (22.97%) were women (33 patients and 142 controls). Of the cases, 72.42% consumed alcohol compared to 50% of the controls. Tobacco also differed greatly between cases (83.09%) and controls (39.80%). None of the polymorphisms were associated to HNSCC risk (Table 1).

Hardy-Weinberg equilibrium showed that the genotypic distributions were in equilibrium for both groups in all polymorphisms studied: *MTHFD1* G1958A (case:  $X^2=1.0876$ ;  $P=0.2970$ , and control:  $X^2=0.0061$ ;  $P=0.9378$ ); *BHMT* G742A (case:  $X^2=0.4320$ ;  $P=0.5110$ , and control:  $X^2=0.7325$ ;  $P=0.3921$ ); *TC2* C776G (case:  $X^2=1.8042$ ;  $P=0.1792$ , and control:  $X^2=0.3262$ ;  $P=0.5679$ ) e *TC2* A67G (case:  $X^2=1.5657$ ;  $P=0.2108$ , and control:  $X^2=0.8329$ ;  $P=0.3614$ ).

For the *MTHFD1* G1958A polymorphism, GG, GA, and AA genotype frequencies were 37.87, 44.85 and 17.28%, respectively, for the cases, and 36.53, 47.96 and 15.51%, respectively, for controls. The variant *MTHFD1* 1958G allele frequencies were 0.60 among the cases and 0.61 among the controls, while the *MTHFD1* 1958A allele frequencies were 0.40 and 0.39 among cases and controls, respectively.

For the *BHMT* G742A polymorphism, GG, GA, and AA genotype frequencies were 43.01, 43.75 and 13.24%, for the cases, and 43.26, 46.33 and 10.41%, respectively, for the controls. The variant *BHMT* 742G allele frequencies were 0.65 among the cases and 0.66 among the controls, while the *BHMT* 742A allele frequencies were 0.35 and 0.34 among cases and controls, respectively.

For the *TC2* C776G polymorphism, CC, CG, and GG genotype frequencies were 37.87, 44.12 and 18.01%, for the cases, and 36.53, 47.96 e 15.51%, respectively, for the controls. The variant *TC2* 776C allele frequencies were 0.60 among the cases and 0.61 among the controls, while the *TC2* 776G allele frequencies were 0.40 and 0.39 among cases and controls, respectively.

For the *TC2* A67G polymorphism, AA, AG, and GG genotype frequencies were 72.29, 23.90 and 3.31%, for the cases, and 71.02, 27.14 and 1.84%, respectively, for the controls. The variant *TC2* 67A allele frequencies were 0.85 among the cases and 0.85 among the controls, while the *TC2* 67G allele frequencies were 0.15 and 0.15 among cases and controls, respectively.

Regarding to polymorphisms and risk factors, we found that tobacco ( $P=0.016$ ) associated to *BHMT* G742A polymorphism modulate head and neck cancer risk (Table 2).

For deleterious alleles, we did not find any relation between allele number and increasead risk for head and neck cancer (one risk: OR:1.02; 95%CI: 0.39-2.66,  $P=0.975$ ; two risk alleles: OR:1.40; 95%CI: 0.55-3.59,  $P=0.481$ ; three risk alleles: OR:0.81; 95%CI: 0.32-2.08,  $P=0.669$ ; four or more risk alleles: OR:1.11; 95%CI: 0.43-2.85,  $P=0.830$ ).

The overall actuarial survival rate was 87.21% in 5 years (Figure 1). The Kaplan-Meier survival curves by genotype are presented in Figure 2, and did not demonstrate any association between polymorphisms and overall survival (*MTHFD1* 1958A,  $P = 0.332$ ; *BHMT* G742A,  $P=0.650$ ; *TC2* C776G,  $P=0.250$ ; *TC2* A67G,  $P=0.807$ ).

## DISCUSSION

Head and neck carcinogenesis is a multifactor process in which environmental etiologic factors cause alterations in oncogenes or tumor suppressor genes. Almost 85% of patients with head and neck cancer are alcohol or tobacco abusers or both. Other causative agents include snuffing or chewing tobacco, ill-fitting dental appliances, chronic candidiasis, viral infection (mainly involving certain HPV types) and poor oral hygiene [1,37,38]. Factors which seem to promote the head and neck field cancerization include environmental exposures to tobacco and alcohol, viral infections especially with human papilloma virus (HPV) subtypes 16 and 18, and deficiency or imbalances in vitamins and micronutrients such as folic acid, vitamins A, C and E, and selenium and zinc [10].

Our results showed that tobacco and male gender were predictors for HNSCC. Tobacco and alcohol are the most important factors predisposing to development of HNSCC [1,38-42], it leads to cells damage and the genetic code [43]. If these alterations in DNA structures are left un-repaired, genetic changes can accumulate, which may result in cell-cycle dysregulation, autonomous growth and development of invasive mechanisms, leading to carcinoma [44]. Furthermore, studies show that male gender remains the most affected by this type of tumor [39,41,45]. However, while the incidence of HNSCC is much higher in males, more and more females are developing HNSCC as women adopt the male pattern of alcohol and tobacco consumption [1].

The understanding that folate metabolism influences both DNA synthesis and methylation has turned environmental and genetic variants into attractive candidates for cancer susceptibility. Genetic variants in genes that encode enzymes involved in one-carbon metabolism are good candidates for studying the impact of both genetic and environmental effects and their interactions on cancer risk. Our study did not find association between studied polymorphisms and risk for HNSCC.

Consequently, deficiency in *MTHFD1* gene may exacerbate the disruption of folate metabolism [46]. Our study did not find any association between *MTHFD1* G1958A polymorphism and head and neck cancer as Kruzyna *et al* (2010) [47] study, that also did not find association between laryngeal cancer and this polymorphism. In other cancer types, the studies are controversy, Liu *et al* (2008) [48], reported that the *MTHFD1* 1958A allele is less frequency in patients with lung cancer by studying a Chinese population with 500 cases and 517 controls. Matakidou *et al* (2007) [49] did not associate lung cancer with this polymorphism in 619 United Kingdom patients with this disease. Difference among these reports could be explained by the difference in study populations, given that UK population is mostly composed by Caucasian, whereas the Liu *et al* (2008) [48] study was restricted to Chinese population. In colorectal cancer, Chen *et al* (2004) [46] and Koushik *et al* (2006) [26] did not relate the polymorphism with the disease. The only study that showed significance between the frequency of methylation in breast cancer and *MTHFD1* G1958A was the study of Li *et al* (2006) [50].

Although not directly involved in folate metabolism, *BHMT* is involved in the metabolism of homocysteine. *BHMT* may play a critical role in the remethylation of homocysteine when the folate-dependent pathway is compromised by either genetic or dietary factors [14,17]. There are no studies in head and neck cancer related to this polymorphism, however in our study, smoking associated to *BHMT* G742A polymorphism increased the risk for this disease. A case-control study of Koushik *et al* (2006)

[26] in 376 colorectal patients and 849 controls observed that 742A polymorphic allele and the GA heterozygous genotype were associated to higher risk of colorectal cancer than those who have the wild-type genotype. The same polymorphic allele was related to a higher survival for those who had HNSCC by Xu *et al* (2008 and 2009) [17,27]. This could be explained by the difference in alimentation and other habits like smoking and etilism.

Cobalamine (Cbl) is an essential nutrient that plays an important role as coenzyme in two metabolic reactions: 1) the conversion of L-methylmalonyl-CoA to succinyl-CoA; 2) the remethylation of homocysteine to methionine. Methyl-Cbl serves as coenzyme for the enzyme methionine synthase (MTR) acting as a carrier for the methyl group donated by 5-methyltetrahydrofolate [51]. Cbl is bound to transcobalamin II, which is required for cellular uptake of this vitamin [19]. The present study showed that *TC2 C776G* and *TC2 A67G* polymorphisms were not associated to HNSCC. In other hand, Hazra *et al* (2007) [52] found that *TC2 C776G* polymorphism is associated with higher risk of colorectal adenoma. Curtin *et al* (2007) [53] also find that 766G allele is associated with colon tumorigenesis. There are no studies in the literature about this association in HNSCC. In relation to *TC2 A67G* polymorphism, there are no studies in the literature in cancer. However, Biselli *et al* (2008) [30] observed an association between this polymorphisms and maternal risk for Down syndrome. Afman *et al* (2002) [28] did not associate *TC2 A67G* and neural tube defects risk.

For deleterious alleles, we did not find any relation between allele number and increasead risk for head and neck cancer. There are no studies in the literature relating deleterious alleles and the polymorphisms evaluated in this study.

In conclusion, the present study suggests that *BHMT G742A* associated to tobacco modulate HNSCC risk. There is no association between *MTHFD1 G1958A* and *TC2 C776G* polymorphisms, risk factors and the disease.

Considering the high prevalence of these polymorphisms in the general population, results from the study will improve the identification of risk factors for disease prevention. However, further investigation of gene-gene interactions in folate metabolism and studies in different populations are needed to investigate polymorphisms and HNSCC risk.

## REFERENCE

- [1] Marcu, LG & Yeoh, E (2009) A review of risk factors and genetic alterations in head and neck carcinogenesis and implications for current and future approaches to treatment. *J Cancer Res Clin Oncol* 135:1303–1314.
- [2] Ragin CCR, Modugno F and Gollin S M (2007) The Epidemiology and Risk Factors of Head and Neck Cancer: a Focus on Human Papillomavirus. *J Dent Res* 86:104.
- [3] Olivieri EHR, da Silva SD, Mendonça FF, Urata YN, Vidal DO, Faria MAM, Nishimoto IN, Rainho CA, Kowalski LP, Rogatto SR (2009) *CYP1A2\*1C*, *CYP2E1\*5B*, and *GSTM1* polymorphisms are predictors of risk and poor outcome in head and neck squamous cell carcinoma patients. *Oral Oncology* - Article In Press.

- [4] Wang Y, Guo W, He Y, Chen Z, Wen D, Zhang X, Wang N, Li Y, Ge H, Zhang J (2007) Association of MTHFR C677T and SHMT1 C1420T with susceptibility to ESCC and GCA in a high incident region of Northern China. *Cancer causes control*, 18:143-52.
- [5] Wang L, Lu J, An J, Shi Q, Spitz MR, Wei O (2007) Polymorphisms of cytosolic serine hydroxyl-methyltransferase and risk of lung cancer: A case—control analysis. *Lung Cancer*; 57:143—151.
- [6] Ma E, Iwasaki M, Junko I, Hamada GS, Nishimoto IN, Carvalho SMT, Motola Jr J, Laginha FM, Tsugane S (2009) Dietary intake of folate, vitamin B6, and vitamin B12, genetic polymorphism of related enzymes, and risk of breast cancer: a case-control study in Brazilian women. *BMC Cancer*; 9:122.
- [7] Linhart HG, Troen A, Bell GW, Cantu E, Chao W, Moran E, Steine E, He T, Jaenisch R (2009) Folate Deficiency Induces Genomic Uracil Misincorporation and Hypomethylation But Does Not Increase DNA Point Mutations. *Gastroenterology*; 136:227–235.
- [8] Garcia-Crespo D, Knock E, Jabado N, Rozen R (2009) Intestinal Neoplasia Induced by Low Dietary Folate Is Associated with Altered Tumor Expression Profiles and Decreased Apoptosis in Mouse Normal Intestine. *The Journal of Nutrition*; 139:488-494.
- [9] Langevin SM, Lin D, Matsuo K, Gao CM, Takezaki T, Stolzenberg-Solomon RZ, Vasavi M, Hasan Q, Taioli E (2009) Review and pooled analysis of studies on MTHFR C677T polymorphism and esophageal cancer. *Toxicology Letters*; 184:73–80.
- [10] Kane, MA (2005) The role of folates in squamous cell carcinoma of the head and neck. *Cancer Detection and Prevention*; 29:46-53.
- [11] Kim, DH (2007) The interactive effect of methyl-group diet and polymorphism of methylenetetrahydrofolate reductase on the risk of colorectal cancer. *Mutat Res*; 622: 14-8.
- [12] Kim, YI (1999) Folate and carcinogenesis: evidence, mechanisms, and implications. *J Nutr Biochem*; 10:66–88.
- [13] Mason JB, Choi SW (2005) Effects of alcohol on folate metabolism: implications for Carcinogenesis. *Alcohol* 35:235–241.
- [14] Xu X, Gammon MD, Wetmur JG, Bradshaw PT, Teitelbaum SL, Neugut AI, Santella RM, Chen J (2008) B-vitamin intake, one-carbon metabolism, and survival in a population based study of women with breast cancer. *Cancer Epidemiol. Biomarkers Prev*; 8:2109-16.
- [15] Kalmbach RD, Choumenkovitch SF, Troen AP, Jacques PF, D’Agostino R, and Selhub J (2008) A 19-Base Pair Deletion Polymorphism in Dihydrofolate Reductase Is Associated with Increased Unmetabolized Folic Acid in Plasma and Decreased Red Blood Cell Folate. *The Journal of Nutrition*; 138:2323–2327.
- [16] Stolzenberg-Solomon RZ, Chang SC, Leitzmann MF, Johnson KA, Johnson C, Buys SS, Hoover RN and Ziegler RG (2006) Folate intake, alcohol use, and postmenopausal breast cancer risk in the Prostate, Lung, Colorectal, and Ovarian Cancer Screening Trial. *Am J Clin Nutr*; 83:895-904.



- [17] Xu X, Gammon MD, Zeisel SH, Lee YL, Wetmur JG, Teitelbaum SL, Bradshaw PT, Neugut AI, Santella RM e Chen J (2008) Choline metabolism and risk of breast cancer in a population-based study. *The FASEB Journal*; 22:2045-2052.
- [18] Hsiung DT, Marsit CJ, Houseman EA, Eddy K, Furniss CS, McClean MD, Kelsey KT (2007) Global DNA methylation level in whole blood as a biomarker in head and neck squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev*; 16:108-14.
- [19] Charasson V, Hillaire-Buys D, Solassol I, Laurand-Quancard A, Pinguet F, Morvan VL, Robert J (2009) Involvement of gene polymorphisms of the folate pathway enzymes in gene expression and anticancer drug sensitivity using the NCI-60 panel as a model. *European Journal of Cancer*.
- [20] Krajcinovic M, Lemieux-Blanchard E, Chiasson S, Primeau M, Costea I, Moghrabi A (2004) Role of polymorphisms in MTHFR and MTHFD1 genes in the outcome of childhood acute lymphoblastic leukemia. *The Pharmacogenomics Journal*; 4:66–72.
- [21] Stevens VL, McCullough ML, Pavluck AL, Talbot JT, Feigelson HS, Thun MJ and Calle EE (2007) Association of Polymorphisms in One-Carbon Metabolism Genes and Postmenopausal Breast Cancer Incidence. *Cancer Epidemiol Biomarkers*; 16:6.
- [22] Weisberg IS, Park E, Ballman KV, Berger P, Nunn M, Suh DS, Breksa 3rd AP, Garrow TA, Rozen R (2003) Investigations of a common genetic variant in betaine-homocysteine methyltransferase (BHMT) in coronary artery disease. *Atherosclerosis*; 167:205–214.
- [23] Morin I, Platt R, Weisberg I, Sabbaghian N, Wu Q, Garrow T.A., Rozen R (2003) Common variant in betaine-homocysteine methyltransferase (BHMT) and risk for spina bifida. *Am J Med Genet*; 119A: 172–176.
- [24] Heil SG, Van der Put NM, Waas ET, den Heijer M, Trijbels FJ, Blom HJ (2001) Is mutated serine hydroxymethyltransferase (SHMT) involved in the etiology of neural tube defects? *Mol Genet Metab*; 73:164-172.
- [25] Zhu, H; Curry, S; Wen, S; Wicker, NJ; Shaw, GM; Lammer, EJ; Yang, W; Jafarov, T; Finnell, RH (2005) Are the Betaine-Homocysteine Methyltransferase (BHMT and BHMT2) Genes Risk Factors for Spina Bifida and Orofacial Clefts? *American J of Medical Genetics*, 135A:274–277.
- [26] Koushik A, Kraft P, Fuchs CS, Hankinson SE, Willett WC, Giovannucci EL, Hunter DJ (2006) Nonsynonymous polymorphisms in genes in the one-carbon metabolism pathway and associations with colorectal cancer. *Cancer Epidemiol Biomarkers Prev*; 15(12):2408-2417.
- [27] Xu X and Chen J (2009) One-carbon metabolism and breast cancer: an epidemiological perspective. *J. Genet. Genomic*; 36:203-214.
- [28] Afman LA, Lievers KJ, Van der Put NM, Trijbels FJ, Blom HJ (2002) Single nucleotide polymorphisms in the transcobalamin gene: relationship with transcobalamin concentrations and risk for neural tube defects. *Eur J Hum Genet*; 10:433–438.
- [29] Miller JW, Ramos MI, Garrod MG, Flynn MA, Green R (2002) Transcobalamin II 775GNC polymorphism and indices of vitamin B12 status in healthy older adults. *Blood*; 15:718–720.

- [30] Biselli JM, Brumati D, Frigeri VF, Zampieri BL, Goloni-Bertollo EM, Pavarino-Bertelli EC (2008) A80G polymorphism of reduced folate carrier 1 (*RFC1*) and C776G polymorphism of transcobalamin 2 (*TC2*) genes in Down's syndrome etiology. *Med J*; 126:329-32.
- [31] <http://www.hemonline.com.br/portarias/rdc153/indexframe.htm>
- [32] Kjaerhein K, Gaard M, Andersen A (1998) The role of alcohol, tobacco, and dietary factors in upper aerogastric tract cancer: a prospective study of 10.900 Norwegian men. *Cancer Causes and Control*; 9:99-108.
- [33] Ahrendt SA, Chown JT, Yang SC, Wu L, Zhang MJ, Jen J, Sidransky D (2000) Alcohol consumption and cigarette smoking increase the frequency of p53 mutations in non-small cell lung cancer. *Cancer Res*; 3155-3159.
- [34] Miller SA, Dikes DD e Polesky HF (1988) A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Research*; 16:1215.
- [35] Hol FA, Van der Put NM, Geurds MP, Heil SG, Trijbels FJ, Hamel BC, Mariman EC, Blom HJ (1998) Molecular genetic analysis of the gene encoding the trifunctional enzyme MTHFD (methylene tetrahydrofolate-dehydrogenase, methenyl tetrahydrofolate-cyclohydro-lase, formyl tetrahydrofolate synthetase) in patients with neural tube defects. *Clin Genet*; 2:119-125.
- [36] Pietrzyk JJ, Bik-Multanowski M (2003) 776C > G polymorphism of the transcobalamin II gene as a risk factor for spina bifida. *Molecular Genetics and Metabolism* 80:364.
- [37] Serefoglou Z, Yapijakis C, Nkenke E, Vairaktaris E (2008) Genetic association of cytokine DNA polymorphisms with head and neck cancer. *Oral Oncology*; 44:1093– 1099.
- [38] Canova, C; Hashibe, M; Simonato, L; Nelis, M; *et al.* (2009) Genetic Associations of 115 Polymorphisms with Cancers of the Upper Aerodigestive Tract across 10 European Countries: The ARCAGE Project. *Cancer Res*; 69:7.
- [39] Galbiatti, ALS; Ruiz, MT; Pinto, DR; Raposo, LS; Maniglia, JV; Pavarino-Bertelli, EC *et al* (2010) A80G polymorphism of reduced folate carrier 1 (*RFC1*) gene and head and neck squamous cell carcinoma etiology in Brazilian Population. *Mol Biol Rep*. [Epub ahead of print]
- [40] Ishiguro S, Sasazuki S, Inoue M, Kurahashia N, Iwasakia M, Tsugane S (2009) Effect of alcohol consumption, cigarette smoking and flushing response on esophageal cancer risk: A population-based cohort study (JPHC study). *Cancer Letters* 275:240–246.
- [41] Argiris, A; Karamouzis, MV; Raben, D; Ferris, RL (2008) Head and neck cancer. *Lancet*; 371:1695-1709.
- [42] Werbrouck, J; De Ruyck, K; Duprez, F; Van Eijkeren, M; Rietzschel, E; Bekaert, S *et al.* (2008) Single-nucleotide polymorphisms in DNA double-strand break repair genes: Association with head and neck cancer and interaction with tobacco use and alcohol consumption. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*; 656:74-81.
- [43] Hoeijmakers, JHJ (2001) Genome maintenance mechanisms for preventing cancer, *Nature*; 411:366–374.

- [44] Scully, C; Field, JK; Tanzawa, H (2000) Genetic aberrations in oral or head and neck squamous cell carcinoma (HNSCC): carcinogen metabolism, DNA repair and cell cycle control. *Oral Oncol.* 36:256–263.
- [45] Parkin, DM; Bray, F; Ferlay, J; Pisani, P (2005) Global cancer statistics, 2002. *CA Cancer J Clin*; 55: 74–108.
- [46] Chen J, Kyte C, Valcin M, Chan W, Wetmur JG, Selhub J, Hunter DJ and MA (2004) Polymorphisms in the one-carbon metabolic pathway, plasma folate levels and colorectal cancer in a prospective study. *J. J Cancer*; 110:617–620.
- [47] Kruszyna, L; Lianeri, M; Rydzanicz, M; *et al* (2010) Polymorphic variants of folate metabolism genes and the risk of laryngeal cancer. *Mol Biol Rep*, 37:241–247.
- [48] Liu, H; Jin, G; Wang, H; Wu, W; Liu, Y; Qian, J (2008) Association of polymorphisms in one-carbon metabolizing genes and lung cancer risk: a case-control study in Chinese population. *Lung Cancer*; 61:21–29.
- [49] Matakidou, A; Galta, R; Rudd, MF; Webb, EL; Bridle, H; Eisen, T; *et al.* (2007) Prognostic significance of folate metabolism polymorphisms for lung cancer. *British Journal of Cancer*, 97:247 – 252.
- [50] Li, SY; Rong, M; Iacopetta, B (2006) Germ-line variants in methyl-group metabolism genes and susceptibility to DNA methylation in human breast cancer. *Oncology Reports*; 21:225.
- [51] Barbosa PR, Stabler SP, Trentin R, Carvalho FR, Luchessi AD, Hirata RDC, Hirata MH, Allen RH, Guerra-Shinohara EM (2008) Evaluation of nutritional and genetic determinants of total homocysteine, methylmalonic acid and S-adenosylmethionine/S-adenosylhomocysteine values in Brazilian childbearing-age women. *Clinica Chimica Acta*; 139–147.
- [52] Hazra, A; Wu, K; Kraft, P; Fuchs, CS; Giovannucci, EL (2007) Twenty-four non-synonymous polymorphisms in the one-carbon metabolic pathway and risk of colorectal adenoma in the Nurses' Health Study. *Carcinogenesis*; 28:1510–1519.
- [53] Curtin, K; Slattery, M; Ulrich, CM; Bigler, J; Levin, TR; Wolff, RK *et al.* (2007) Genetic polymorphisms in one-carbon metabolism: associations with CpG island methylator phenotype (CIMP) in colon cancer and the modifying effects of diet. *Carcinogenesis*; 28:1672–1679.

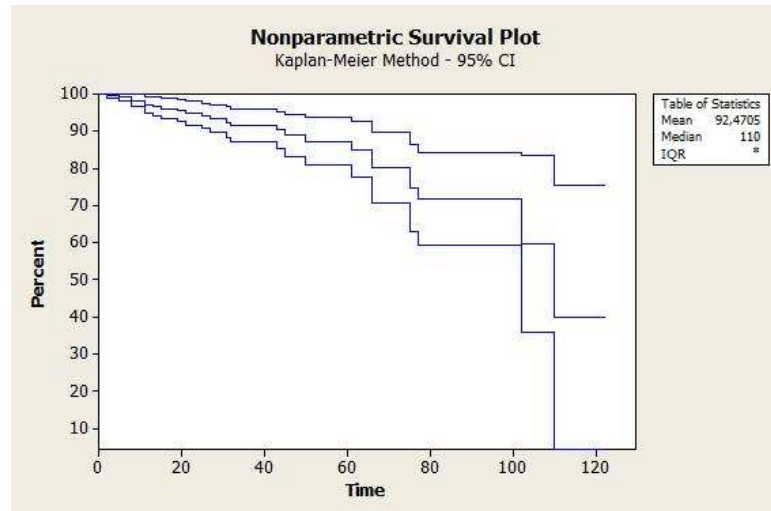
**Table1.** Distribution in odds ratio (OR) of the gender, polymorphisms and risk factors between head and neck squamous cell carcinoma patients and controls

Variables	Patients N (%)	Controls N (%)	OR (95%CI)	P value
<b>Tobacco consumption</b>				
Non-smokers	41 (17.08)	267 (58.81)	Reference	Referência
Smokers	199 (82.92)	187 (41.19)	3.89 (2.45-6.19)	<b>P&lt;0.05</b>
<b>Alcohol consumption</b>				
Alcohol non-consumers	67 (27.92)	230 (50.66)	Reference	Referência
Alcohol consumers	173 (72.08)	224 (49.34)	1.58 (0.99-2.51)	<b>P&lt;0.05</b>
<b>Gender</b>				
Female	29 (12.08)	129 (28.41)	Reference	Referência
Male	211 (87.92)	325 (71.59)	1.64 (0.94-2.84)	P=0.08
<b>Age</b>				
<42 years	8 (3.33)	177 (38.99)	Reference	Referência
42-51 years	49 (20.42)	170 (37.44)	5.36 (2.59-11.09)	<b>P&lt;0.05</b>
52-63 years	99 (41.25)	51 (11.23)	29.02 (13.61-61.88)	<b>P&lt;0.05</b>
>64 years	84 (35)	56 (12.34)	25.26 (11.87-53.75)	<b>P&lt;0.05</b>
<b>MTHFD1 G1958A genotypes</b>				
GG	86 (35.83)	161 (35.46)	Reference	Referência
GA	110 (45.84)	221 (48.68)		
AA	44 (18.33)	72 (15.86)	1.39 (0.91-2.12)	P=0.12
<b>BHMT G742A genotypes</b>				
GG	101 (42.08)	188 (41.41)	Reference	Referência
GA	109 (24)	215 (47.35)		
AA	30 (33.92)	51 (11.24)	1.02 (0.68-1.53)	P=0.20
<b>TC2 C776G genotypes</b>				
CC	85 (35.42)	156 (34.36)	Reference	Referência
CG	109 (45.42)	227 (50)		
GG	46 (19.16)	71 (15.64)	0.76 (0.50-1.16)	P=0.91
<b>TC2 A67G genotypes</b>				
AA	178 (74.17)	320 (70.48)	Reference	Referência
AG	55 (22.92)	126 (27.75)		
GG	7 (2.91)	8 (1.77)	1.21 (0.78-1.87)	P=0.40

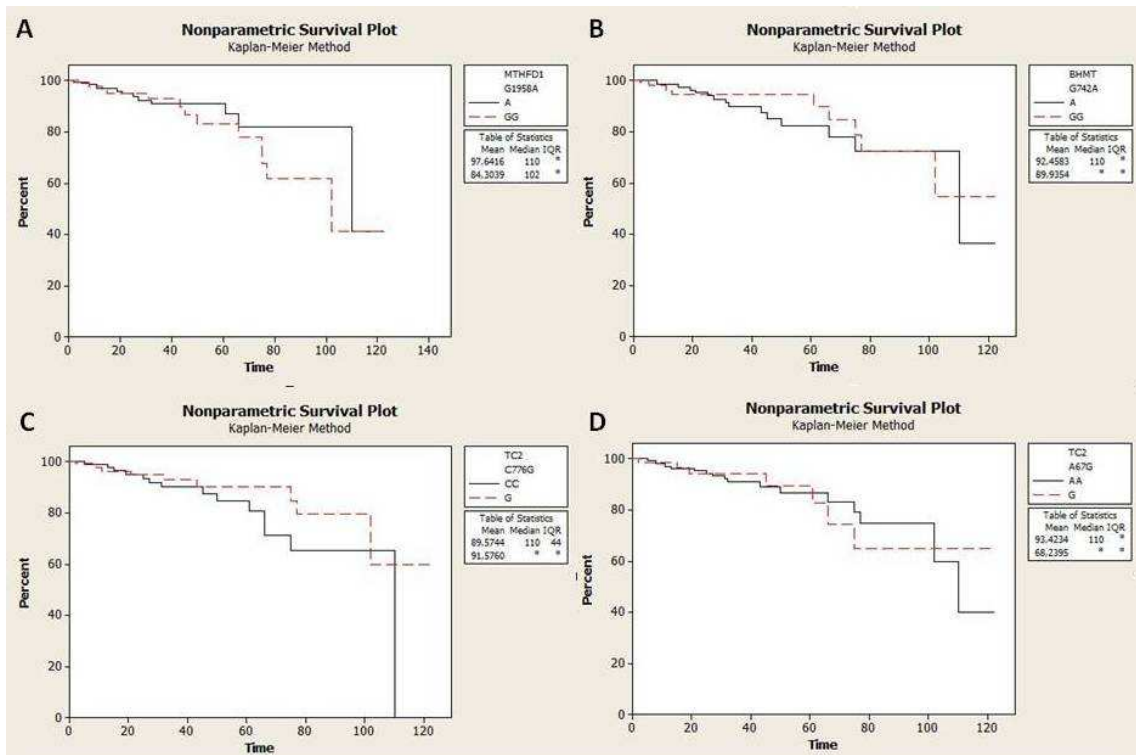
**Table 2.** Odds ratio of head and neck cancer related to *MTHFD1*, *BHMT* and *TC2* genotypes by age, gender, tobacco and alcohol consumption

Risk Factors	OR (95%CI) – P value			
	<i>MTHFD1</i> G1958A*	<i>BHMT</i> G742A*	<i>TC2</i> C776G*	<i>TC2</i> A67G*
<b>Tobacco consumption</b>				
Yes	1.69 (1.01-2.80) <b>P = 0.044</b>	1.07 (0.66-1.74) P = 0.790	0.68 (0.41-1.14) P = 0.144	1.24 (0.73-2.08) P = 0.427
No	1.00 (0.45-2.20) P = 0.997	0.89 (0.41-1.90) P = 0.756	0.78 (0.35-1.73) P = 0.536	1.40 (0.59-3.34) P = 0.441
<b>Alcohol consumption</b>				
Yes	1.81 (1.05-3.14) <b>P = 0.033</b>	1.28 (0.76-2.15) P = 0.353	0.70 (0.40-1.22) P = 0.212	0.99 (0.56-1.74) P = 0.972
No	1.01 (0.51-2.01) P = 0.968	0.66 (0.34-1.29) P = 0.223	0.82 (0.42-1.63) P = 0.576	1.62 (0.79-3.33) P = 0.185
<b>Gender</b>				
Female	1.34 (0.47-3.85) P = 0.581	0.40 (0.15-1.07) P = 0.068	1.01 (0.34-2.97) P = 0.989	0.79 (0.27-2.33) P = 0.676
Male	1.37 (0.86-2.21) P = 0.188	1.19 (0.75-1.88) P = 0.455	0.75 (0.47-1.21) P = 0.239	1.34 (0.82-2.20) P = 0.248
<b>Age</b>				
<42 years	0.37 (0.09-1.52) P = 0.167	0.85 (0.21-3.45) P = 0.825	1.34 (0.30-5.88) P = 0.699	0.93 (0.21-4.12) P = 0.923
42-51 years	1.84 (0.85-3.97) P = 0.120	1.49 (0.74-3.02) P = 0.263	1.29 (0.62-2.68) P = 0.503	2.02 (0.96-4.24) P = 0.063
52-63 years	2.35 (0.95-5.81) P = 0.065	1.72 (0.69-4.32) P = 0.247	0.35 (0.13-0.95) <b>P = 0.039</b>	0.37 (0.14-0.98) <b>P = 0.046</b>
>64 years	1.47 (0.66-3.28) P = 0.347	0.55 (0.25-1.22) P = 0.140	0.40 (0.17-0.95) <b>P = 0.039</b>	1.94 (0.79-4.76) P = 0.148

\* *MTHFD1* G1958A Reference: GG wild type genotype; *BHMT* G742A Reference: GG wild type genotype; *TC2* C776G Reference: CC wild type genotype; *TC2* A67G Reference: AA wild type genotype.



**Figure1.** Overall survival rates by Kaplan-Meier method.



**Figure2.** Nonparametric survival plot (Kaplan Meier) from patients with head and neck cancer. (A: *MTHFD1* G1958A; B: *BHMT* G742A; C: *TC2* C776G and D: *TC2* A67G).

### 3 CONCLUSÕES

### 3. Conclusões

1. Os polimorfismos estudados *MTHFD1* G1958A, *BHMT* G742A, *TC2* C776G e *TC2* A67G não estão associados ao desenvolvimento do câncer de cabeça e pescoço.
2. Os genótipos *MTHFD1* 1958GA ou 1958AA associados ao tabagismo e etilismo e os genótipos *BHMT* 742GA ou 742AA associados ao tabagismo, modulam o risco de desenvolver carcinoma espinocelular de cabeça e pescoço.
3. Os genótipos *MTHFD1* 1958GA ou 1958AA apresenta-se mais frequentes em indivíduos com estadios tumorais 3 e 4 e estão associados a uma sobrevida menor em relação ao genótipo *MTHFD1* 1958GG.



## *4 REFERÊNCIAS BIBLIOGRÁFICAS*

#### 4. Referências Bibliográficas

1. Ragin CCR, Modugno F and Gollin S M. The Epidemiology and Risk Factors of Head and Neck Cancer: a Focus on Human Papillomavirus. *J Dent Res* 2007; (86):104.
2. Marcu, LG & Yeoh, E. A review of risk factors and genetic alterations in head and neck carcinogenesis and implications for current and future approaches to treatment. *J Cancer Res Clin Oncol* 2009; (135):1303–1314.
3. INCA – Instituto Nacional de Câncer: [www.inca.gov.br](http://www.inca.gov.br), 2010.
4. Olivieri EHR, da Silva SD, Mendonça FF, Urata YN, Vidal DO, Faria MAM, Nishimoto IN, Rainho CA, Kowalski LP, Rogatto SR. CYP1A2\*1C, CYP2E1\*5B, and GSTM1 polymorphisms are predictors of risk and poor outcome in head and neck squamous cell carcinoma patients. *Oral Oncology* 2009 - Article In Press.
5. Vioque J, Barber X, Bolumar F, Porta M, Santibáñez M, Heral MG, Moreno-Osset E. Esophageal cancer risk by type of alcohol drinking and smoking: a case-control study in Spain. *BMC Cancer* 2008; (8):221.
6. Argiris A, Karamouzis MV, Raben D, Ferris RL. Head and neck cancer. *Lancet* 2008; (371):1695-1709.
7. Sapkota A, Hsu CC, Zaridze D, Shangina O, Szeszenia-Dabrowska N, Mates D, Fabianova E, Rudnai P, Janout V, Holcatova I, Brennan P, et. al. Dietary risk factors for squamous cell carcinoma of the upper aerodigestive tract in central and eastern Europe. *Cancer Causes Control* 2008; (19):1161–1170.
8. Serefoglou Z, Yapijakis C, Nkenke E, Vairaktaris E. Genetic association of cytokine DNA polymorphisms with head and neck cancer. *Oral Oncology* 2008; (44):1093– 1099.

9. Werbrouck J, De Ruyck K, Duprez F, Van Eijkeren M, Rietzschel E, Bekaert S, Vral A, De Neve W and Thierens H. Single-nucleotide polymorphisms in DNA double-strand break repair genes: Association with head and neck cancer and interaction with tobacco use and alcohol consumption. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis* 2008; v.656, Issues 1-2, (30):74-81.
10. Yadav SS, Ruwali M, Shah PP, Mathur N, Singh RL, Pant MC, Parmar D. Association of poor metabolizers of cytochrome P450 2C19 with head and neck cancer and poor treatment response. *Mutation Research* 2008; (644):31–37.
11. Ishiguro S, Sasazuki S, Inoue M, Kurahashia N, Iwasakia M, Tsugane S. Effect of alcohol consumption, cigarette smoking and flushing response on esophageal cancer risk: A population-based cohort study (JPHC study). *Cancer Letters* 2009; (275):240–246.
12. Ruwali M, Khan AJ, Shah PP, Singh AP, Pant MC, Parmar D. Cytochrome P450 2E1 and Head and Neck Cancer: Interaction With Genetic and Environmental Risk Factors. *Environmental and Molecular Mutagenesis* 2009; (50):473-482.
13. Boonkitticharoen V, Kulapaditharom B, Leopairut J, Kraiphibul P, Larbcharoensub N, Cheewaruangroj W, Chintrakarn C, Pochanukul L. Vascular Endothelial Growth Factor A and Proliferation Marker in Prediction of Lymph Node Metastasis in Oral and Pharyngeal Squamous Cell Carcinoma. *Arch Otolaryngol Head Neck Surg* 2008; 134(12):1305-1311.
14. Kane, MA. The role of folates in squamous cell carcinoma of the head and neck. *Cancer Detection and Prevention* 2005; (29):46-53.

15. Eleftheriadou A, Chalastras T, Ferekidou E, Yiotakis I, Kyriou L, Tzagarakis M. Association between squamous cell carcinoma of the head and neck and serum folate and homocysteine. *Anticancer Res* 2006; (26):2345-2348.
16. Lo AK, Lo KW, Tsao SW, Wong HL, Hui JW, To KF, Hayward DS, Chui YL, Lau YL, Takada K, Huang DP. Epstein-Barr virus infection alters cellular signal cascades in human nasopharyngeal epithelial cells. *Neoplasia* 2006; (3):173-180.
17. Hennessey PT, Westra WH, Califano JA. Human papillomavirus and head and neck squamous cell carcinoma: recent evidence and clinical implications. *Dent Res* 2009; (4):300-306.
18. Langevin SM, Lin D, Matsuo K, Gao CM, Takezaki T, Stolzenberg-Solomon RZ, Vasavi M, Hasan Q, Taioli E. Review and pooled analysis of studies on MTHFR C677T polymorphism and esophageal cancer. *Toxicology Letters* 2009; (184):73-80.
19. Kim, YI. Folate and carcinogenesis: evidence, mechanisms, and implications. *J Nutr Biochem* 1999. (10):66-88.
20. Mason JB, Choi SW. Effects of alcohol on folate metabolism: implications for Carcinogenesis. *Alcohol* 2005; (38):235-241.
21. Xu X, Gammon MD, Wetmur JG, Rao M, Gaudet MM, Teitelbaum SL, Britton JA, Neugut AI, Santella RM, *et al.* A functional 19-base pair deletion polymorphism of dihydrofolate reductase (DHFR) and risk of breast cancer in multivitamin users. *Am J Clin Nutr* 2007; (85):1098-1102.
22. Kalmbach RD, Choumenkovitch SF, Troen AP, Jacques PF, D'Agostino R, and Selhub J. A 19-Base Pair Deletion Polymorphism in Dihydrofolate Reductase Is Associated with Increased Unmetabolized Folic Acid in Plasma and Decreased Red Blood Cell Folate. *The Journal of Nutrition* 2008; (138): 2323-2327.
23. Suzuki T, Matsuo K, Hirose K, Hiraki A, Kawase T, Watanabe M, *et al.* One-carbon metabolism-related gene polymorphisms and risk of breast cancer. *Carcinogenesis* 2008; (29):356-362.

24. Alshatwi AA. Breast cancer risk, dietary intake, and methylenetetrahydrofolate reductase (MTHFR) single nucleotide polymorphisms. *Food and Chemical Toxicology* 2010; (48):1881–1885.
25. Johansson M, Relton C, Ueland PM, *et al.* Serum B Vitamin Levels and Risk of Lung Cancer. *JAMA* 2010; (23):2377-2385.
26. Garcia-Crespo D, Knock E, Jabado N, Rozen R. Intestinal Neoplasia Induced by Low Dietary Folate Is Associated with Altered Tumor Expression Profiles and Decreased Apoptosis in Mouse Normal Intestine. *The Journal of Nutrition*, 2009; (139):488-494.
27. Linhart HG, Troen A, Bell GW, Cantu E, Chao W, Moran E, Steine E, He T, Jaenisch R. Folate Deficiency Induces Genomic Uracil Misincorporation and Hypomethylation But Does Not Increase DNA Point Mutations. *Gastroenterology* 2009; (136):227–235.
28. Bravi F, Polesel J, Bosetti C, Talamini R, Negri E, Dal Maso L, Serraino D, La Vecchia C. Dietary intake of selected micronutrients and the risk of pancreatic cancer: an Italian case–control study. *Annals of Oncology Advance Access* 2010, June 7.
29. Oaks BM, Dodd KW, Meinhold CL, Jiao L, Church TR and Stolzenberg-Solomon RZ. Folate intake, post-folic acid grain fortification, and pancreatic cancer risk in the Prostate, Lung, Colorectal, and Ovarian Cancer Screening Trial. *Am J Clin Nutr* 2010; (91):449–55.
30. Gibson TM, Weinstein SJ, Mayne ST, Pfeiffer RM, Selhub J, Taylor PR, Virtamo J, Albanes D, Stolzenberg-Solomon RZ. A prospective study of one-

- carbon metabolism biomarkers and risk of renal cell carcinoma. *Cancer Causes Control* 2010; (21):1061–1069.
- 31.** Hsiung DT, Marsit CJ, Houseman EA, Eddy K, Furniss CS, McClean MD, Kelsey KT. Global DNA methylation level in whole blood as a biomarker in head and neck squamous cell carcinoma. : *Cancer Epidemiol Biomarkers Prev.* 2007; (16):108-14.
- 32.** Mu LN, Cao W, Zhang ZF, Yu SZ, Jiang QW, You NC, et al. Polymorphisms of 5,10-methylenetetrahydrofolate reductase (MTHFR), fruit and vegetable intake, and the risk of stomach. *Canc. Biomark.* 2007; (12): 61-75.
- 33.** Ouerhani S, Oliveira E, Marrakchi R, Ben Slama MR, Sfaxi M, Ayed M, et al. Methylenetetrahydrofolate reductase and methionine synthase polymorphisms and risk of bladder cancer in a Tunisian population. *Cancer Genet Cytogenet* 2007; (176):48-53.
- 34.** Pande M, Chen J, Amos CI, Lynch PM, Broaddus R, Frazier ML. Influence of Methylenetetrahydrofolate Reductase Gene Polymorphisms C677T and A1298C on Age-Associated Risk for Colorectal Cancer in a Caucasian Lynch Syndrome Population. *Cancer Epidemiol Biomarkers Prev* 2007; (16):1753-1759.
- 35.** Ott N, Geddert H, Sarbia M. Polymorphisms in methionine synthase (A2756G) and cystathionine beta-synthase (844ins68) and susceptibility to carcinomas of the upper gastrointestinal tract. *J Cancer Res Clin Oncol* 2008; (134):405-410.
- 36.** Johanning GL, Heimbürger DC, Piyathilake CJ. DNA methylation and diet in cancer. *J Nutr* 2002; (132):3814-3818.

37. Bailey LB. Folate, Methyl-Related Nutrients, Alcohol, and the MTHFR 677C-T Polymorphism Affect Cancer Risk: Intake Recommendation. *Am Soc Nutr Sci* 2003; (133):3748-3753.
38. Charasson V, Hillaire-Buys D, Solassol I, Laurand-Quancard A, Pinguet F, Morvan VL, Robert J. Involvement of gene polymorphisms of the folate pathway enzymes in gene expression and anticancer drug sensitivity using the NCI-60 panel as a model. *European Journal of Câncer* 2009, Article in Press.
39. Kraunz KS, Hsiung D, McClean MD, Liu M, Osanyingbemi J, Nelson HH, Kelsey KT. Dietary folate is associated with *p16<sup>INK4A</sup>* methylation in head and neck squamous cell carcinoma. *Int. J. Cancer* 2006; (119):1553–1557.
40. Ma E, Iwasaki M, Junko I, Hamada GS, Nishimoto IN, Carvalho SMT, Motola Jr J, Laginha FM, Tsugane S. Dietary intake of folate, vitamin B6, and vitamin B12, genetic polymorphism of related enzymes, and risk of breast cancer: a case-control study in Brazilian women. *BMC Cancer* 2009; (9):122.
41. D'Alessio AC, Szyf M. Epigenetic tête-à-tête: the bilateral relationship between chromatin modifications and DNA methylation. *Biochem Cell Biol* 2006; (84):463-76.
42. Ehrlich M. Expression of various genes is controlled by DNA methylation during mammalian development. *J Cell Biochem* 2003; (88): 899-910.
43. Tuck-Muller CM, Narayan A, Tsien F, Smeets DF, Sawyer J, Fiala ES, et al. DNA hypomethylation and unusual chromosome instability in cell lines from ICF syndrome patients. *Cytogenet Cell Genet* 2000; (89): 121-128.
44. Jones PA, Baylin SB. The fundamental role of epigenetic events in cancer. *Nat Rev Genet.* 2002; (3):415-428.

45. Sciandrello G, Caradonna F, Mauro M, Barbata G. Arsenic-induced DNA hypomethylation affects chromosomal instability in mammalian cells. *Carcinogenesis* 2004; (25):413-507
46. Xu X, Chen J. One-carbon metabolism and breast cancer: an epidemiological perspective. *J. Genet. Genomics* 2009; (36):203-214.
47. Miller JW, Ramos MI, Garrod MG, Flynn MA, Green R. Transcobalamin II 775GNC polymorphism and indices of vitamin B12 status in healthy older adults. *Blood* 2002; (15):718–720.
48. Monsen AL, Refsum H, Markestad T, Ueland PM. Cobalamin status and its biochemical markers methylmalonic acid and homocysteine in different age groups from 4 days to 19 years. *Clin Chem* 2003; (49):2067-2075.
49. Hubner RA and Houlston RS. Folate and colorectal cancer prevention. *British Journal of Cancer* 2009; (100):233–239.
50. Kim DH. The interactive effect of methyl-group diet and polymorphism of methylenetetrahydrofolate reductase on the risk of colorectal cancer. *Mutat Res* 2007; (622):14-18.
51. Zhang W, Press OA, Haiman CA, Yang DY, Gordon MA, Fazzone W, et al. Association of methylenetetrahydrofolate reductase gene polymorphisms and sex-specific survival in patients with metastatic colon cancer. *J Clin Oncol* 2007; (25):3726-3731.
52. Jonge R, Tissing WJE, Hooijberg JH, Jansen G, Kaspers GJL, Lindemans J, Peters GJ, Pieters R. Polymorphisms in folate-related genes and risk of pediatric acute lymphoblastic leukemia. *Blood* 2009; (113):2284-2289.



53. Haddad R, Mendes MA, Hoehr NF, Eberlin MN. Amino acid quantitation in aqueous matrices via trap and release membrane introduction mass spectrometry: homocysteine in human plasma. *Analyst* 2001; (126):1212–1215.
54. Födinger M, Dierkes J, Skoupy S, Röhrer C, Hagen W, Puttinger H, et al. Effect of Glutamate Carboxypeptidase II and Reduced Folate Carrier polymorphisms on folate and total homocysteine concentrations in dialysis patients. *J Am Soc Nephrol* 2003; (14):1314–1319.
55. Stevens VL, McCullough ML, Pavluck AL, Talbot JT, Feigelson HS, Thun MJ and Calle EE. Association of Polymorphisms in One-Carbon Metabolism Genes and Postmenopausal Breast Cancer Incidence. *Cancer Epidemiol Biomarkers* 2007; (6):16.
56. Chen J, Kyte C, Valcin M, Chan W, Wetmur JG, Selhub J, Hunter DJ and MA Polymorphisms in the one-carbon metabolic pathway, plasma folate levels and colorectal cancer in a prospective study. *J. J Cancer* 2004; (110):617–620.
57. Krajcinovic M, Lemieux-Blanchard E, Chiasson S, Primeau M, Costea I, Moghrabi A. Role of polymorphisms in MTHFR and MTHFD1 genes in the outcome of childhood acute lymphoblastic leukemia. *The Pharmacogenomics Journal* 2004; (4):66–72.
58. Weisberg IS, Park E, Ballman KV, Berger P, Nunn M, Suh DS, Breksa 3rd AP, Garrow TA, Rozen R. Investigations of a common genetic variant in betaine-homocysteine methyltransferase (BHMT) in coronary artery disease. *Atherosclerosis* 2003; (167):205–214.
59. Xu X, Gammon MD, Zeisel SH, Lee YL, Wetmur JG, Teitelbaum SL, Bradshaw PT, Neugut AI, Santella RM e Chen J. Choline metabolism and risk

- of breast cancer in a population-based study. *The FASEB Journal* 2008, (22):2045-2052.
60. Koushik A, Kraft P, Fuchs CS, Hankinson SE, Willett WC, Giovannucci EL, Hunter DJ. Nonsynonymous polymorphisms in genes in the one-carbon metabolism pathway and associations with colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 2006; (12):2408-2417.
61. Afman LA, Lievers KJ, Van der Put NM, Trijbels FJ, Blom HJ. Single nucleotide polymorphisms in the transcobalamin gene: relationship with transcobalamin concentrations and risk for neural tube defects. *Eur J Hum Genet* 2002; (10):433-438.
62. Barbosa PR, Stabler SP, Trentin R, Carvalho FR, Luchessi AD, Hirata RDC, Hirata MH, Allen RH, Guerra-Shinohara EM. Evaluation of nutritional and genetic determinants of total homocysteine, methylmalonic acid and S-adenosylmethionine/S-adenosylhomocysteine values in Brazilian childbearing-age women. *Clinica Chimica Acta* 2008; (388):139-147.
63. Quadros EV, Regec AL, Khan KM, Quadros E, Rothenberg SP. Transcobalamin II synthesized in the intestinal villi facilitates transfer of cobalamin to the portal blood. *Am J Physiol.* 1999; (27):161-166.
64. Namour F, Oliver JR, Abdelmouttaalebi I, Adjalla C, Debard R, Salvat C et al. Transcobalamin codon 259 polymorphism in HT-29 and Caco-2-cells and in Caucasians: relation to transcobalamin and homocysteine concentration in blood. *Blood* 2001; (97):1092-1098.
65. Biselli JM, Brumati D, Frigeri VF, Zampieri BL, Goloni-Bertollo EM, Pavarino-Bertelli EC. A80G polymorphism of reduced folate carrier 1 (*RFC1*) and C776G

polymorphism of transcobalamin 2 (*TC2*) genes in Down's syndrome etiology.  
Med J 2008; (6):329-332.

- 66.** Semmeler A, Linnebank M, Krex D, Götz A, Moskau S, Ziegler A, Simon M.  
Polymorphisms of Homocysteine Metabolism Are Associated with Intracranial  
Aneurysms. Cerebrovasc D 2008; (26):425–429.

*5 OUTRAS PRODUÇÕES CIENTÍFICAS*

**Artigo.**

**Título:** Clinical and epidemiological characteristics of patients of the head and neck surgery department in a university hospital of northwest state of São Paulo

**Periódico:** Head and Neck, submetido para publicação.

**Title: Clinical and epidemiological characteristics of patients of the head and neck surgery department in a university hospital of northwest state of São Paulo**

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**ABSTRACT**

**BACKGROUND:** Head and neck cancer is located in upper aerodigestive tract, and accounts for nearly 650,000 new cases worldwide. Squamous cell carcinoma is the most frequent histological type, and the main risk factors are tobacco and alcoholism.

**PATIENTS AND METHODS:** A total of 995 patients from a head and neck surgery department were evaluated. The variables analyzed included: age, gender, skin color, tobacco and alcohol consumption, primary site, histological tumor type, stage, treatment and number of deaths. **RESULTS:** This disease was more frequent among men (79.70%), smokers (75.15%) and alcohol consumers (58.25%). The most represented results were oral cavity (29.65%) and larynx (24.12%) for primary site, squamous cell carcinoma (84.92%) for histological type, surgery (29.04%) and radiotherapy (14.19%) for patient treatment. **CONCLUSION:** Tumors in patients treated by the head and neck surgery department occur mainly in males, tabagists and etilists, with the oral cavity and larynx having the highest incidence.

**Keywords:** Epidemiology, cancer, head and neck surgery department, alcohol and tobacco.

## Introduction

Head and neck cancer is not a specific entity, but rather a broad category of diverse tumor types arising from various anatomic structures including the craniofacial bones, soft tissues, salivary glands, skin, and mucosal membranes.<sup>1</sup>

Although the head and neck surgery department treats patients with malignant tumors of the upper aero digestive tract, skin and thyroid, the term "head and neck cancer" is used only for the group of neoplasms located in the upper aero digestive tract, with approximately 40% occurring in the oral cavity, 15% in the pharynx, 25% in larynx and 20% in other anatomic sites.<sup>2,3</sup> Approximately 95% of these tumors have squamous cell carcinoma as the primary histological type.<sup>4</sup>

Head and neck cancer is the fifth most common cancer worldwide among all neoplasias.<sup>4</sup> The overall survival rate for this cancer is variable, depending on the primary site and disease stage. For oral cavity cancer, the overall survival rate is 50% over five years.<sup>5</sup> For other sites (pharynx and larynx), the rate is above 50% for early stage disease (T1-T2, N0) and generally below 50% in advanced stage (T3-T4, N0, T3-T4, N+, or any T, N2-N3).<sup>6</sup> In 2008, 6,214 deaths were observed in Brazil as a result of this disease.<sup>7</sup>

Oral cavity cancer, the most representative site of the disease, has a high frequency in Southeast Asia and India, due to the habit within such regions of chewing tobacco leaf and betel nut, a stimulant commonly used among Indians.<sup>8</sup> In the United States, about 21,000 new cases of oral cancer are diagnosed each year and it is estimated that more than 650,000 new cases of head and neck cancer are diagnosed each year worldwide, two-thirds of them in developed countries.<sup>6</sup>



The estimated new cases of oral cavity cancer for 2010 is 14,120 and 4,120 for Brazil and Sao Paulo State, respectively.<sup>7</sup> The preliminary search conducted by our group in a period of five years, in a reference hospital in the northwestern state of Sao Paulo showed 427 patients diagnosed with head and neck cancer.<sup>10</sup>

This tumor type occurs mainly in male subjects, and its occurrence increases with age.<sup>11</sup> In the last decade, there has been a significant increase in this cancer in younger individuals, possibly due to increased infections by human papilloma virus (HPV).<sup>6,12,13</sup>

The development of head and neck cancer is the result of the interaction of both environmental factors and genetic inheritance, and is therefore, multifactorial. Tobacco use associated with alcohol consumption is a well-established risk factor for the head and neck cancer.<sup>14</sup> Alcohol can act as a solvent for some tobacco carcinogens, increasing cellular uptake of these. According Maruri and Forastiere (2008),<sup>6</sup> the consumption of tobacco associated with alcohol consumption increases head and neck cancer risk by a rate of 40 times.

A significant proportion of head and neck tumors is a result of infection by some HPV types. These virus affect cells and produce viral oncoproteins (E6 and E7) that promote tumor progression by inactivating the product of some tumor suppressor genes such as Tp53 (tumor protein 53) and pRb (retinoblastoma tumor suppressor gene)<sup>13</sup>. Other factors that may contribute to head and neck carcinogenesis include diet, with risk reduced by fruit and vegetable consumption, and increased risk by<sup>15</sup> inadequate oral hygiene, leading to chronic infections by bacteria responsible for the pathogenesis of this tumor type, and<sup>16</sup> body mass, which can modulate toxin and carcinogenic metabolism.<sup>17</sup>

Occupational activity also appears to be associated with development of head and neck cancer. The study by Conway et al. (2010)<sup>18</sup> showed that manual occupational activities, low income, low occupational-social class, low educational attainment and unemployment correlate with increased risk for disease development. The individuals who work in rural activities are in constant exposure to sunlight and in contact with carcinogenic substances that contribute to the development of oral cavity cancer.<sup>19</sup>

Skin cancer is also associated with excessive exposure to solar radiation and occurs more frequently in the portions of the body exposed to the sun (head, neck and limbs). Also influencing the appearance of these lesions are factors such as age, sex, ethnicity, smoking, alcohol abuse, geographical distribution, old scars, persistent physical aggression and exposure to radioactives.<sup>20</sup>

Thyroid gland tumors, with their own unique characteristics, but also treated in the head and neck surgery department, have an origin related to iodine deficiency, external beam radiotherapy in childhood and adolescence, exposure to ionizing radiation and pre-existing thyroid disease.<sup>21</sup>

This study aimed to describe the socio-demographic aspects and clinical-pathology of patients in head and neck surgery department treated at a university hospital in the northwestern state of Sao Paulo, from January 2000 to May 2010.

### **Patients and methods**

We retrospectively evaluated medical records of 1,351 cancer patients treated at the Otolaryngology and Head and Neck department of a university hospital in the northwestern state of Sao Paulo, from January 2000 to May 2010.

The variables analyzed were age, gender, skin color, tobacco and alcohol consumption, primary site, histological type, staging, treatment, death and occupational therapy of patients with the upper aero digestive tract, skin and thyroid cancer. Individuals who smoked more than 100 cigarettes in their lifetime were considered smokers, and individuals who consumed at least four drinks per week were considered alcohol drinkers.<sup>22,23</sup>

Tumors of the upper aero digestive tract were classified according to the anatomical site in the oral cavity, pharynx, larynx, nasal cavity, salivary glands and unknown primary site. Skin tumors and thyroid cases were also included.

Clinical staging of patients was performed according to the International Union Against Cancer based on the classification of malignant tumors (TNM).<sup>24</sup> According to these standards, for the classification stage, T represents the tumor size, and tumors classified as Tx and T0 indicate primary indefinite tumor and no signs of primary tumor, respectively. N1, N2 and N3 indicate the existence of lymph nodes and N0 the absence of them. Tumors classified as Nx indicate undetermined lymph node status. Metastasis is represented by M1 and M0 for absence. In cases of failure to diagnose the presence or absence of metastasis, tumors were classified as Mx.

The occupations of patients were classified in such sectors as agriculture, construction, domestic service, driver, commercial, administrative, surveillance, metalwork, tapestries and aesthetics.

Data were analyzed using descriptive statistics, using the software Excel (version 2007).

## **Results**

We analyzed records of 1,351 patients, of whom it was possible to obtain the most comprehensive information in hospital records of 995 cases, except for tumor staging and treatment, which were obtained for 785 and 909 cases, respectively.

A substantial majority of the cases were males (79.68%) and the mean age of patients was 60.48 years. Of the total patients, 747 were smokers (75.15%), 579 were etilists (58.25%) and 547 (54.00%) were both. For skin color classification, the subjects were divided into white and nonwhite, according to medical records, and was found a 90.04% frequency of patients with white skin. The oral cavity was the primary site of occurrence (29.65%), followed by the larynx and pharynx (24.12% and 18.29%, respectively). The thyroid gland tumors corresponded to 5.43%, while skin tumors accounted for 6.83% of patients. In 110 patients, it was not possible to identify the primary site (Table 1). The predominant histological type was squamous cell carcinoma, representing 84.92% of cases, followed by basal cell carcinoma (6.03%) and papillary carcinoma (5.22%). Other types of malignant tumors such as adenocarcinoma, melanoma, sarcoma, chondrosarcoma, fibrosarcoma and follicular carcinoma accounted for 3.08% (Table 2).

The tumor stage (TNM) in relation to primary sites and the main methods of treatment made by patients are described in Tables 3 and 4, respectively.

Regarding the occupation of the patients, the main activity was related to agriculture represented by 207 patients (20.82%) (Table 5).

## **Discussion**

In the present study, the mean age of patients, regardless of gender, was 60.48 years, similar to that observed in Brazilian<sup>25</sup> and American<sup>26</sup> populations. Although the

frequency of patients with head and neck cancer is higher in individuals with advanced age, an increasing number of cases, especially for oral cavity and oropharynx cancer in young people has been observed and associated with HPV 16 infection,<sup>13</sup> an etiological agent related to carcinogenesis of this tumor type.<sup>12</sup>

Head and neck cancer affects mostly males<sup>10, 27-29</sup>. Likewise, in the present study, 79.68% of individuals affected by this type of cancer were males. Despite the low incidence of malignancies in women, an increase in number of cases is expected as a result of increased tobacco and alcohol consumption in the female population.<sup>30</sup>

In the present study, a prevalence of white individuals (90.04%) was observed, similar to findings in a study conducted recently in southern states of Brazil<sup>31</sup> and Midwest United States.<sup>32</sup> However, Hayat et al (2007)<sup>33</sup> observed a higher prevalence of head and neck cancer in African-American descendents from different areas of the United States. The ethnic differences in the distribution found in patients with head and neck cancer in different studies are mainly due to population composition where the research was done. Our study was conducted in northwestern São Paulo State, predominantly colonized by Europeans, comprising a high percentage of white-skinned people in the population.

Tobacco consumption was 75.15%; alcohol consumption was 58.25%; and both were 54.00% among the patients studied. The association between alcohol and tobacco with head and neck cancer is well established and has been reported in several studies<sup>14, 27, 34</sup>. An association between experiencing passive smoking for over 15 years and the development of head and neck cancer has been reported, independent of alcohol consumption.<sup>35</sup>

Squamous cell carcinoma was the most common histological type, representing 84.82% of cases, with a frequency close to that observed in the literature, which is approximately 90%<sup>5</sup>. In relation to primary site of tumor, oral cavity was the most representative (29.65%), followed by larynx (24.12%) and pharynx (18.29%). The incidence of oral cancer worldwide is highest compared to other anatomical sites and is more common in individuals with low income, low occupational social class, low educational attainment.<sup>36</sup> Although it was not possible to obtain all information in our study, most patients treated in the hospital were from public health system, which assists low-income patients.

Surgical procedure and radiotherapy were performed in 29.04% and 14.19% of patients, respectively. The use of surgical practice followed by radiotherapy is a common practice in the treatment of head and neck squamous cell carcinoma, especially in early stages of the disease (I or II) with a high percentage of cure.<sup>37</sup> In this study, both treatment procedures were used in 29.92% of cases. In advanced stages (III or IV), chemotherapy is usually used in conjunction with other forms of treatment, promoting, especially in conjunction with radiotherapy, increased locoregional control in many cases.<sup>37</sup> Among the patients analyzed in this study, 13.86% underwent surgical, radiotherapy and chemotherapy procedures.

Patients with skin tumors in our study underwent surgery, which has cure rates above 95% when treated early and properly.<sup>20</sup>

Regarding thyroid gland tumors, our results indicate the prevalence of surgery associated with iodine therapy as an optional treatment. The use of less conservative surgery and the exploration of lymph nodes, as well as the use of adjuvant iodine

therapy seems to determine a more favorable prognosis for patients with thyroid cancer.<sup>38</sup>

In the analysis of the tumor stage, a high proportion of tumors classified as T3 and T4 were observed. These data reveal a significant number of patients diagnosed in advanced stages of disease and demonstrated the difficulty in obtaining an early diagnosis, since symptoms rarely appear in the early stages. According to the literature, on average, 40% of patients with oral cancer are diagnosed in advanced stages.<sup>39</sup>

In Brazil, some studies reported less than 50% of patients receive an early diagnosis<sup>40,41</sup>. Nearly two-thirds of patients with head and neck cancer have an advanced stage of the disease, usually involving regional lymph nodes. The incidence of distant metastases is relatively small in malignant tumors of head and neck cancer compared with other regions.<sup>42</sup> Approximately one-third of patients of this study had lymph node involvement and only 1% of metastasis.

During the study period, 265 patients died. The high mortality rate for this tumor type remains virtually constant over past decades.<sup>11</sup> Nevertheless Zigon et al (2010)<sup>28</sup> observed an increase in the relative rate of five-year survival for head and neck cancers study in the European population. In another study in the Brazilian population, we observed a significant increase in survival rate over five years for oral and oropharyngeal cancer, ranging from 28.7% in patients treated in the 1950s to 43.2% in the 1990s.<sup>41</sup>

In relation to occupational activities performed by study subjects, the most frequent were those related to agriculture (20.82%) and construction (19.01%), which is consistent with Conway et al.<sup>36</sup> who showed a correlation between higher rates of head

and neck cancer and individuals who practice manual occupational activities and have a low occupational social class.

Sartor et al.<sup>43</sup> also showed an association between laryngeal cancer and exposure to respirable-free crystalline silica. In this study, we found a risk twice as high for exposed individuals, such as those working in construction, compared to unexposed individuals.

Recent research in molecular biology has broadened our understanding of the etiology of these tumors. The combination of prognostic factors and molecular parameters could be beneficial for patients with the application of new therapeutic strategies. Such advances in studies of molecular markers may also improve the diagnosis in early stages of the disease, which would not be possible by traditional clinical methods<sup>27,44</sup>.

## **Conclusion**

Malignant tumors that affect patients treated by head and neck surgery department of a university hospital in northwestern São Paulo State are more frequent in subjects from regions with low socioeconomic development, and limited access to education. The incidence of this disease in this specific population affects mostly male patients in their sixties, smokers and etilists, with the oral cavity and larynx being the regions most affected. The high rate of patients with stage III and IV indicates a late demand in treatment centers, which reflects the need for prevention education campaigns for early diagnosis of the disease.



## References

1. Pai SI & Westra WH. Molecular Pathology of Head and Neck Cancer; Implications for Diagnosis, Prognosis, and Treatment. *Annu. Rev. Pathol. Mech Dis.* 2009; 4:49-70.
2. Dobrossy L. Epidemiology of head and neck cancer: magnitude of the problem. *Cancer and Metastasis Rev* 2005; 24:9-17.
3. Lee KJ. *Essential Otolaryngology: Head & Neck Surgery*. The McGraw-Hill Companies, 2003; 8.
4. Marcu LG, Yeoh E. A review of risk factors and genetic alterations in head and neck carcinogenesis and implications for current and future approaches to treatment. *J Cancer Res Clin Oncol.* 2009;135:1303-14.
5. Walker DM, Boey G, McDonald LA. The pathology of oral cancer. *Pathology.* 2003;35:376-83
6. Marur S, Forastiere AA. Head and neck cancer: changing epidemiology, diagnosis, and treatment. *Mayo Clin Proc.* 2008;83:489-501.
7. Home Page: Instituto Nacional do Câncer. Disponível em <http://www.inca.gov.br>. [acessado em 27 de junho de 2010].
8. Kanavos P. The rising burden of cancer in the developing world. *Ann Oncol.* 2006; 17.
9. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. *CA Cancer J Clin.* 2009;59.
10. Alvarenga Lde M, Ruiz MT, Pavarino-Bertelli EC, Ruback MJ, Maniglia JV, Goloni-Bertollo M. Epidemiologic evaluation of head and neck patients in a university hospital of Northwestern São Paulo State. *Braz J Otorhinolaryngol.* 2008; 74:68-73.

11. Crozier E, Sumer BD. Head and neck cancer. *Med Clin North Am.* 2010, 94.
12. Gillison ML, D'Souza G, Westra W, Sugar E, Xiao W, Begum S, Viscidi R. Distinct risk factor profiles for human papillomavirus type 16-positive and human papillomavirus type 16-negative head and neck cancers. *J Natl Cancer Inst.* 2008;100:407-20
13. Hennessey PT, Westra WH, Califano JA. Human papillomavirus and head and neck squamous cell carcinoma: recent evidence and clinical implications. *J Dent Res.* 2009; 88:300-6.
14. Boffetta P, Hashibe M. Alcohol and cancer. *Lancet Oncol.* 2006;7:149-56.
15. Pavia M, Pileggi C, Nobile CG, Angelillo IF. Association between fruit and vegetable consumption and oral cancer: a meta-analysis of observational studies. *Am J Clin Nutr.* 2006;83:1126-34.
16. Guha N, Boffetta P, Wunsch Filho V, Eluf Neto J, Shangina O, Zaridze D, Curado MP, Koifman S, Matos E, Menezes A, Szeszenia-Dabrowska N, Fernandez L, Mates D, Daudt AW, Lissowska J, Dikshit R, Brennan P. Oral health and risk of squamous cell carcinoma of the head and neck and esophagus: results of two multicentric case-control studies. *Am J Epidemiol* 2007; 166:1159–1173.
17. Peters ES, LUCKETT BG, Applebaum KM, Marsit CJ, McClean MD, Kelsey KT. Dairy products, leanness, and head and neck squamous cell carcinoma. *Head Neck* 2008; 30:1193–1205.
18. Conway DI, McMahon AD, Smith K, Black R, Robertson G, Devine J, McKinney PA. Components of socioeconomic risk associated with head and neck cancer: A population-based case-control study in Scotland. *British Journal of Oral and Maxillofacial Surgery* 48 2010 11–17.

19. Santos LCO, Cangussu MCT, Batista OM, Santos JP. Oral Cancer: Population Sample of the State of Alagoas at a Reference Hospital. *Braz J Otorhinolaryng.* 2009; 75.
20. Dergham AP, Muraro CC, Ramos EA, Mesquita LAF, Collaço LM, Collaço LM. Distribution of diagnosis of neoplastic and pre neoplastic skin lesions at Evangelical Hospital in Curitiba. *An bras Dermatol* 2004; 79(5):555-9
21. Golbert L, Wajner SM, Rocha AP, Maia AL, Gross JL. Carcinoma Diferenciado de Tireóide: Avaliação Inicial e Acompanhamento. *Arq Bras Endocrinol Metab* 2005;49
22. Ahrendt SA, Chown JT, Yang SC, Wu L, Zhang MJ, Jen J, Sidransky D. Alcohol consumption and cigarette smoking increase the frequency of p53 mutations in non small cell lung cancer. *Cancer Res* 2000; 60:3155–3159
23. Kjaerhein K, Gaard M, Andersen A. The role of alcohol, tobacco, and dietary factors in upper aerogastric tract cancer: a prospective study of 10,900 Norwegian men. *Cancer Causes Control* 1998; 9:99–108
24. Sobin LH & Wittelind CH. International union against cancer: TNM classification of malignant tumors. 6th ed. New York: Wiley, 2000.
25. Vilela LD, Allison PJ. An investigation of the role of sense of coherence in predicting survival among Brazilians with head and neck cancer. *Oral Oncol.* 2010;46:531-5.
26. Potash AE, Karnell LH, Christensen AJ, Vander Weg MW, Funk GF. Continued alcohol use in patients with head and neck cancer. *Head Neck.* 2010;32:905-12.
27. Galbiatti AL, Ruiz MT, Chicote-Biselli PM, Raposo LS, Maniglia JV, Pavarino-Bertelli EC, Goloni-Bertollo EM. 5-Methyltetrahydrofolate-homocysteine

methyltransferase gene polymorphism (MTR) and risk of head and neck cancer. *Braz J Med Biol Res.* 2010;43:445-50.

28. Zigon G, Berrino F, Gatta G, Sanchez MJ, van Dijk B, Van Eycken E, Francisci S. Prognoses for head and neck cancers in Europe diagnosed in 1995–1999: a population - based study. *Annals of Oncology Advance Access*, 2010.

29. St Guily JL, Borget I, Vainchtock A, Rémy V, Takizawa C. Head and neck cancers in France: an analysis of the hospital medical information system (PMSI) database. *Head Neck Oncol.* 2010;2:22.

30. La Vecchia C, Lucchini F, Negri E, Levi F. Trends in oral cancer mortality in Europe. *Oral Oncol.* 2004; 40:433-9.

31. Silver HJ, de Campos Graf Guimaraes C, Pedruzzi P, Badia M, Spuldaro de Carvalho A, Oliveira BV, Ramos GH, Dietrich MS, Pietrobon R. Predictors of functional decline in locally advanced head and neck cancer patients from south Brazil. *Head Neck.* 2010;32:1217-25.

32. Shuman AG, Entezami P, Chernin AS, Wallace NE, Taylor JM, Hogikyan ND. Demographics and efficacy of head and neck cancer screening. *Otolaryngol Head Neck Surg.* 2010;14.

33. Hayat MJ, Howlader N, Reichman ME, Edwards BK. Cancer statistics 2007. *Oncologist* 2007; 12:20-37.

34. Hashibe M, Boffetta P, Zaridze D, Shangina O, Szeszenia-Dabrowska N, Mates D, Fabiánová E, Rudnai P, Brennan P. Contribution of tobacco and alcohol to the high rates of squamous cell carcinoma of the supraglottis and glottis in Central Europe. *Am J Epidemiol.* 2007;165:814-20

35. Lee YC, Boffetta P, Sturgis EM, Wei Q, Zhang ZF, Muscat J, Lazarus P, Matos E, Hayes RB, Winn DM, Zaridze D, Wünsch-Filho V, Eluf-Neto J, Koifman S, Mates D, Curado MP, Menezes A, Fernandez L, Daudt AW, Szeszenia-Dabrowska N, Fabianova E, Rudnai P, Ferro G, Berthiller J, Brennan P, Hashibe M. Involuntary smoking and head and neck cancer risk: pooled analysis in the international head and neck cancer epidemiology consortium. *Cancer Epidemiol Biomarkers Prev* 2008; 17:1974–1981.
36. Conway DI, Peticrew M, Marlborough H, Berthiller J, Hashibe M, Macpherson LM. Socioeconomic inequalities and oral cancer risk: a systematic review and meta-analysis of case-control studies. *Int J Cancer* 2008; 122:2811–2819.
37. Argiris A, Karamouzis MV, Raben D, Ferris RL. Head and neck cancer. *Lancet*. 2008; 371:1695-709.
38. Rouxel A, Hejblum G, Bernier MO, Boelle PY, Menegaux F, Mansour G, et al. Prognostic factors associated with the survival of patients developing loco-regional recurrences of differentiated thyroid carcinomas. *J Clin Endocrinol Metab* 2004;89(11):5362-8.
39. Rogers SN, Pabla R, McSorley A, Lowe D, Brown JS, Vaughan ED. An assessment of deprivation as a factor in the delays in presentation, diagnosis and treatment in patients with oral and oropharyngeal squamous cell carcinoma. *Oral Oncol*. 2007; 43:648-55.
40. Wünsch-Filho V. The epidemiology of oral and pharynx cancer in Brazil. *Oral Oncol*. 2002; 38:737–46.

41. Carvalho AL, Ikeda MK, Magrin J, Kowalski LP. Trends of oral and oropharyngeal cancer survival over five decades in 3267 patients treated in a single institution. *Oral Oncol.* 2004, 40.
42. Ferlito A, Shahab AR, Silverc CE, Rinaldo A. Incidence and Sites of Distant Metastases from Head and Neck Cancer. *ORL* 2001;63:202–7.
43. Sartor SG, Eluf-Neto J, Travier N, Wunsch Filho V, Arcuri AS, Kowalski LP et al. Riscos ocupacionais para o câncer de Laringe: um estudo caso-controle. *Cad Saúde Pública*, 2007, 23:1473-81.
44. Ruiz MT, Bertelli EP, Maniglia JV, Ruback MJC, Goloni-Bertollo EM. Epidemiologia e biomarcadores de em câncer de cabeça e pescoço. *Arq Ciênc Saúde.* 2006;13:34-8

**Table 1.** Distribution of cases according to demographic characteristics and sites of tumor.

<b>Variables</b>	<b>Number of patients (%)</b>
<b>Gender</b>	
Male	793 (79.70)
Female	202 (20.30)
<b>Skin color</b>	
White	895 (90.04)
Non-white	99 (9.96)
<b>Tobacco use</b>	
Tobacco users	747 (75.15)
Non tobacco users	247 (24.85)
<b>Alcohol use</b>	
Etilists	579 (58.25)
Non-etilists	415 (41.75)
<b>Tobacco and alcohol use</b>	547 (54.00)
<b>Tumor sites</b>	
Oral cavity	295 (29.65)
Larynx	240 (24.12)
Pharynx	182 (18.29)
Skin	68 (6.83)
Thyroid	54 (5.43)
Nasal cavity	16 (1.61)
Other sites	30 (3.01)
Unknown primary site	110 (11.06)
<b>DP = standard deviation</b>	

**Table 2.** Most frequent histological types in patients attending a head and neck surgery department.

<b>Histological types</b>	<b>Number of patients (%)</b>
Squamous cell carcinoma	845 (84.92)
Basal cell carcinoma	60 (6.03)
Papillary carcinoma	52 (5.22)
Other	38 (3.83)

**Table 3.** Case distribution by clinical-histopathological characteristics.

Category	Tumoral Staging					
	T1 e T2 n (%)	T3 e T4 n (%)	N0 n (%)	N1, N2 e N3 n (%)	M0 n (%)	M1 n (%)
Oral Cavity	169 (40.14)	102 (31.19)	202 (37.75)	69 (25.46)	249 (33.73)	2 (28.57)
Larynx	105 (24.94)	114 (34.86)	160 (29.90)	64 (23.62)	208 (28.18)	-----
Pharynx	66 (15.68)	98 (27.97)	89 (16.60)	78 (28.78)	141 (19.10)	2 (28.57)
Skin	39 (9.26)	1 (0.31)	41 (7.66)	-----	41 (5.55)	-----
Thyroid	24 (5.70)	7 (2.14)	23 (4.29)	5 (1.85)	28 (3.79)	-----
Nasal Cavity	5 (1.19)	3 (0.92)	7 (1.13)	1 (0.37)	8 (1.08)	-----
Other Sites	13 (3.09)	2 (0.61)	13 (2.49)	3 (1.11)	15 (2.03)	-----
Unknown Site	0 (0)	0 (0)	0 (0)	51 (18.82)	48 (6.54)	3 (42.86)

**Table 4.** Treatment forms of patients attending the head and neck surgery department.

Treatment	Number of Patients (%)
Surgery	264 (29.04)
Radiotherapy	129 (14.19)
Chemotherapy	18 (1.98)
Iodotherapy	1 (0.11)
Surgery and Iodotherapy	23 (2.53)
Surgery and Radiotherapy	272 (29.92)
Surgery and Chemotherapy	17 (1.87)
Radiotherapy and Chemotherapy	59 (6.49)
Surgery, Radiotherapy and Chemotherapy	126 (13.86)



**Table 5.** Occupation of patients attending the head and neck surgery department.

Ocupacional activities	Number of patients (%)
Farming	207 (20.82)
Civil construction	189 (19.01)
Domestic services	175 (17.60)
Driver	89 (8.95)
Commercial	82 (8.25)
Administrative	43 (4.32)
Vigilance	26 (2.61)
Metallurgy	21 (2.11)
Tapestry	9 (0.90)
Aesthetics	5 (0.50)
Other*	149 (14.89)

\* Less than 1% in each profession

***6 ANEXOS***

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## FACULDADE DE MEDICINA DE SÃO JOSÉ DO RIO PRETO

Autarquia Estadual - Lei n° 8899 de 27/09/94  
(Reconhecida pelo Decreto Federal n° 74.179 de 14/06/74)

### COMITÊ DE ÉTICA EM PESQUISA

O Comitê de Ética em Pesquisa da Faculdade de Medicina de São José do Rio Preto tomou ciência e aprovou a ampliação da metodologia datada de 11 de novembro de 2005, referente ao protocolo n.º 5566/2005 sob a responsabilidade de Maurício José Cabral Ruback, com o título "Câncer de cabeça e pescoço: um levantamento epidemiológico do Hospital de Base/FAMERP de São José do Rio Preto".

São José do Rio Preto, 19 de dezembro de 2005.

Prof. Dr. José Paulo Cipullo  
Vice- Coordenador do CEP/FAMERP



## FACULDADE DE MEDICINA DE SÃO JOSÉ DO RIO PRETO

Autarquia Estadual - Lei n.º 8899 de 27/09/94  
(Reconhecida pelo Decreto Federal n.º 74.179 de 14/06/74)

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### COMITÊ DE ÉTICA EM PESQUISA

O Comitê de Ética em Pesquisa em Seres Humanos da Faculdade de Medicina de São José do Rio Preto tomou ciência e autorizou a **dispensa do Termo de Consentimento Livre e Esclarecido conforme solicitação datado de 03/07/2008**, referente ao protocolo n.º **5566/2005** sob a responsabilidade de **Maurício José Cabral Ruback**, com o título "Câncer de cabeça e pescoço: um levantamento epidemiológico do Hospital de Base/FAMERP de São José do Rio Preto".

São José do Rio Preto, 08 de julho de 2008.

Prof. Dr. Antonio Carlos Pires  
Coordenador do OEP/FAMERP